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amino acid sequences which are at least about 50% identical to the amino acid sequence shown in SEQ ID NO: 2 and; the amino acid sequence shown in SEQ ID NO: 2;

5 Binding between the test compound and the elongase HSEL01-like protein peptide is detected. A test compound which binds to the elongase HSEL01-like protein polypeptide is thereby identified as a potential agent for decreasing extracellular matrix degradation. The agent can work by decreasing the activity of the elongase HSEL01-like protein.

10 Another embodiment of the invention is a method of screening for agents which decrease extracellular matrix degradation. A test compound is contacted with a polynucleotide encoding a elongase HSEL01-like protein polypeptide, wherein the polynucleotide comprises a nucleotide sequence selected from the group consisting of:

15 nucleotide sequences which are at least about 50% identical to the nucleotide sequence shown in SEQ ID NOS: 1, the nucleotide sequence shown in SEQ ID NOS: 1,

20 nucleotide sequences which are at least about 50% identical to the nucleotide sequence shown in SEQ ID NOS: 5, the nucleotide sequence shown in SEQ ID NOS: 5,

25 nucleotide sequences which are at least about 50% identical to the nucleotide sequence shown in SEQ ID NOS: 6 and,

Binding of the test compound to the polynucleotide is detected. A test compound which binds to the polynucleotide is identified as a potential agent for decreasing extracellular matrix degradation. The agent can work by decreasing the amount of the

elongase HSEL01-like protein through interacting with the elongase HSEL01-like protein mRNA.

Another embodiment of the invention is a method of screening for agents which regulate extracellular matrix degradation. A test compound is contacted with a elongase HSEL01-like protein polypeptide comprising an amino acid sequence selected from the group consisting of:

amino acid sequences which are at least about 50% identical to

5 the amino acid sequence shown in SEQ ID NO: 2 and; the amino acid sequence shown in SEQ ID NO: 2;

A elongase HSEL01-like protein activity of the polypeptide is detected. A test compound which increases elongase HSEL01-like protein activity of the polypeptide relative to elongase HSEL01-like protein activity in the absence of the test compound is thereby identified as a potential agent for increasing extracellular matrix degradation. A test compound which decreases elongase HSEL01-like protein activity of the polypeptide relative to elongase HSEL01-like protein activity in the absence of the test compound is thereby identified as a potential agent for decreasing extracellular matrix degradation.

Even another embodiment of the invention is a method of screening for agents which decrease extracellular matrix degradation. A test compound is contacted with a elongase HSEL01-like protein product of a polynucleotide which comprises a nucleotide sequence selected from the group consisting of:

25 nucleotide sequences which are at least about 50% identical to the nucleotide sequence shown in SEQ ID NO: 1,

the nucleotide sequence shown in SEQ ID NO: 1,

30 nucleotide sequences which are at least about 50% identical to

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the nucleotide sequence shown in SEQ ID NO: 5,
the nucleotide sequence shown in SEQ ID NO: 5,
nucleotide sequences which are at least about 50% identical to
the nucleotide sequence shown in SEQ ID NO 6 and,
5 the nucleotide sequence shown in SEQ ID NO: 6,

Binding of the test compound to the elongase HSEL01-like protein product is detected. A test compound which binds to the elongase HSEL01-like protein product is thereby identified as a potential agent for decreasing extracellular matrix degradation.

Still another embodiment of the invention is a method of reducing extracellular matrix degradation. A cell is contacted with a reagent which specifically binds to a polynucleotide encoding a elongase HSEL01-like protein polypeptide or the product encoded by the polynucleotide, wherein the polynucleotide comprises a nucleotide sequence selected from the group consisting of:

nucleotide sequences which are at least about 50% identical to
the nucleotide sequence shown in SEQ ID NOS: 1,

the nucleotide sequence shown in SEQ ID NO: 1,

nucleotide sequences which are at least about 50% identical to
the nucleotide sequence shown in SEQ ID NO: 5,

the nucleotide sequence shown in SEQ ID NO: 5,
nucleotide sequences which are at least about 50% identical to
the nucleotide sequence shown in SEQ ID NO: 6 and,
20 the nucleotide sequence shown in SEQ ID NO: 6,

nucleotide sequences which are at least about 50% identical to
the nucleotide sequence shown in SEQ ID NO: 6 and,
the nucleotide sequence shown in SEQ ID NO: 6,

30 Elongase HSEL01-like protein activity in the cell is thereby decreased.

The invention thus provides a human elongase HSEL01-like protein that can be used to identify test compounds that may act, for example, as activators or inhibitors at the enzyme's active site. Human elongase HSEL01-like protein and fragments thereof also are useful in raising specific antibodies that can block the enzyme and effectively reduce its activity.

BRIEF DESCRIPTION OF THE DRAWINGS

Fig. 1 shows the DNA-sequence encoding a elongase HSEL01-like protein Polypeptide (SEQ ID NO: 1).

5 Fig. 2 shows the amino acid sequence deduced from the DNA-sequence of Fig. 1 (SEQ ID NO: 2).

10 Fig. 3 shows the amino acid sequence of the protein identified by trembl|AF292387|AF292387_1 (SEQ ID NO: 3).

15 Fig. 4 shows the DNA-sequence encoding a elongase HSEL01-like protein Polypeptide (SEQ ID NO: 4).

20 Fig. 5 shows the DNA-sequence encoding a elongase HSEL01-like protein Polypeptide (SEQ ID NO: 5).

25 Fig. 6 shows the DNA-sequence encoding a elongase HSEL01-like protein Polypeptide (SEQ ID NO: 6).

30 Fig. 7 shows the amino acid sequence of a protein identified by trembl|AF034504|AF034504_1 (SEQ ID NO: 7).

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Fig. 8 shows the amino acid sequence of a elongase HSELO1-like protein Polypeptide (SEQ ID NO: 8).

Fig. 9 shows the amino acid sequence of a elongase HSELO1-like protein Polypeptide (SEQ ID NO: 9).

Fig. 10 shows the amino acid sequence of a elongase HSELO1-like protein Polypeptide (SEQ ID NO: 10).

Fig. 11 shows the BLASTP - alignment of 397_protein (SEQ ID NO: 2) against trembl|AF292387|AF292387_1 (SEQ ID NO: 3).

Fig. 12 shows the BLASTP - alignment of 397_protein (SEQ ID NO: 2) against trembl|AF054504|AF054504_1 (SEQ ID NO: 7).

Fig. 13 shows the BLASTP - alignment of 397_protein (SEQ ID NO: 2) against trembl|AF170907|AF170907_1 (SEQ ID NO: 8).

Fig. 14 shows the BLASTP - alignment of 397_protein (SEQ ID NO: 2) against trembl|AF170908|AF170908_1 (SEQ ID NO: 9).

Fig. 15 shows the BLASTP - alignment of 397_protein (SEQ ID NO: 2) against swiss|P40319|SUR4_YEAST (SEQ ID NO: 10).

Fig. 16 shows the HMM/PFAM - alignment of 397_protein (SEQ ID NO: 2) against pfam|pfam|GNS1_SUR4.

Fig. 17 shows the BLOCK search result.

Fig. 18 shows the Genewise output.

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Fig. 19 shows the TM HMM result

Fig. 20 shows the BLASTP - alignment of 397_protein against tremblnew|AF277094|AF277094_1

DETAILED DESCRIPTION OF THE INVENTION

The invention relates to an isolated polynucleotide from the group consisting of:

- 10 a) a polynucleotide encoding a elongase HSELO1-like protein polypeptide comprising an amino acid sequence selected from the group consisting of:
 - 15 10 a) a polynucleotide which are at least about 50% identical to the amino acid sequence shown in SEQ ID NO: 2 and; the amino acid sequence shown in SEQ ID NO: 3.
 - 20 15 b) a polynucleotide comprising the sequence of SEQ ID NOS: 1, 5 or 6.
 - 25 20 c) a polynucleotide which hybridizes under stringent conditions to a polynucleotide specified in (a) and (b) and encodes a elongase HSELO1-like protein polypeptide;
 - 30 25 d) a polynucleotide the sequence of which deviates from the polynucleotide sequences specified in (a) to (c) due to the degeneration of the genetic code and encodes a elongase HSELO1-like protein polypeptide; and
 - 35 30 e) a polynucleotide which represents a fragment, derivative or allelic variation of a polynucleotide sequence specified in (a) to (d) and encodes a elongase HSELO1-like protein polypeptide.

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Furthermore, it has been discovered by the present applicant that a novel elongase HSEL01-like protein, particularly a human elongase HSEL01-like protein can be used in therapeutic methods to treat cancer, diabetes, CNS disorder, asthma, metabolic disease or COPD.

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Human elongase HSEL01-like protein comprises the amino acid sequence shown in SEQ ID NO: 2. Coding sequences for human elongase HSEL01-like protein are shown in SEQ ID NOS: 1, 5, and 6. The human elongase HSEL01-like protein gene is located on chromosome 10. A related EST (SEQ ID NO: 4) is expressed in adenocarcinoma.

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By sequence similarity, PFAM, and BLOCK search, SEQ ID NO: 2 is a human homolog/ortholog of mouse CIG30 and belongs GNS1/SUR4 family (GNS1 and SUR4 are synonymous names for ELO2 and ELO3 respectively, which are believed to function in very long chain fatty acid elongation). GNS1/SUR4 family proteins are evolutionary related-integral membrane proteins. Although its exact function has not yet clearly been established, CIG30 is believed to be a long chain polyunsaturated fatty acid elongation enzyme. Human elongase HSEL01-like protein are all related to their mouse homolog or ortholog (SSC1, SSC2, and CIG30, products of Ssc1, Ssc2, and Cig30, respectively). Mouse Cig30, Ssc1 and Ssc2 are newly discovered genes. Experimental findings indicate that these proteins are involved in synthesis of very long chain fatty acids and sphingolipid. Alignments are provided in FIGS. 1-6.

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Human elongase HSEL01-like protein is believed to be useful in therapeutic methods to treat disorders such as cancer, diabetes, and CNS disorders. Human elongase HSEL01-like protein also can be used to screen for human elongase HSEL01-like protein activators and inhibitors.

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Variations in percent identity can be due, for example, to amino acid substitutions, insertions, or deletions. Amino acid substitutions are defined as one for one amino acid replacements. They are conservative in nature when the substituted amino acid has similar structural and/or chemical properties. Examples of conservative replacements are substitution of a leucine with an isoleucine or valine, an aspartate with a glutamate, or a threonine with a serine.

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Amino acid insertions or deletions are changes to or within an amino acid sequence. They typically fall in the range of about 1 to 5 amino acids. Guidance in determining

Polypeptides

Human elongase HSEL01-like protein polypeptides according to the invention comprise at least 6, 10, 15, 20, 25, 50, 75, 100, 125, 150, 175, 200, 225, 250, 275, or 296 contiguous amino acids selected from the amino acid sequence shown in SEQ ID NO: 2 or a biologically active variant thereof, as defined below. An elongase HSEL01-like protein polypeptide of the invention therefore can be a portion of an elongase HSEL01-like protein, a full-length elongase HSEL01-like protein, or a fusion protein comprising all or a portion of an elongase HSEL01-like protein.

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Biologically Active Variants

Human elongase HSEL01-like protein polypeptide variants that are biologically active, e.g., retain an elongase activity, also are elongase HSEL01-like protein polypeptides. Preferably, naturally or non-naturally occurring elongase HSEL01-like protein polypeptide variants have amino acid sequences which are at least about 50, 55, 60, 65, or 70, preferably about 75, 80, 85, 90, 95, 96, 98, or 99% identical to the amino acid sequence shown in SEQ ID NO: 2 or a fragment thereof. Percent identity between a putative elongase HSEL01-like protein polypeptide variant and an amino acid sequence of SEQ ID NO: 2 is determined using the Blast2 alignment program (Blossum62, Expect 10, standard genetic codes).

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Variations in percent identity can be due, for example, to amino acid substitutions, insertions, or deletions. Amino acid substitutions are defined as one for one amino acid replacements. They are conservative in nature when the substituted amino acid has similar structural and/or chemical properties. Examples of conservative replacements are substitution of a leucine with an isoleucine or valine, an aspartate with a glutamate, or a threonine with a serine.

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Amino acid insertions or deletions are changes to or within an amino acid sequence. They typically fall in the range of about 1 to 5 amino acids. Guidance in determining

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which amino acid residues can be substituted, inserted, or deleted without abolishing biological or immunological activity of an elongase HSEL01-like protein polypeptide can be found using computer programs well known in the art, such as DNASTAR software. Whether an amino acid change results in a biologically active elongase HSEL01-like protein polypeptide can readily be determined by assaying for enzyme activity, as described for example, in Ghioni et al., Biochim Biophys Acta 1999 Feb 25;1437(2):170-81; Tocher, Lipids 1993 Apr;28(4):267-72; or Chang et al., J Nutr 1992 Nov;122(11):2074-80.

Fusion Proteins

10 Fusion proteins are useful for generating antibodies against elongase HSEL01-like protein polypeptide amino acid sequences and for use in various assay systems. For example, fusion proteins can be used to identify proteins that interact with portions of an elongase HSEL01-like protein polypeptide. Protein affinity chromatography or library-based assays for protein-protein interactions, such as the yeast two-hybrid or phage display systems, can be used for this purpose. Such methods are well known in the art and also can be used as drug screens.

15 An elongase HSEL01-like protein polypeptide fusion protein comprises two polypeptide segments fused together by means of a peptide bond. The first polypeptide segment comprises at least 6, 10, 15, 20, 25, 50, 75, 100, 125, 150, 175, 200, 225, 250, 275, or 296 contiguous amino acids of SEQ ID NO: 2 or of a biologically active variant, such as those described above. The first polypeptide segment also can comprise full-length elongase HSEL01-like protein.

20 The second polypeptide segment can be a full-length protein or a protein fragment. Proteins commonly used in fusion protein construction include β -galactosidase, β -glucuronidase, green fluorescent protein (GFP), autofluorescent proteins, including blue fluorescent protein (BFP), glutathione-S-transferase (GST), luciferase, horse-radish peroxidase (HRP), and chloramphenicol acetyltransferase (CAT). Addition-

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ally, epitope tags are used in fusion protein constructions, including histidine (His) tags, FLAG tags, influenza hemagglutinin (HA) tags, Myc tags, VSV-G tags, and thioredoxin (Trx) tags. Other fusion constructions can include maltose binding protein (MBP), S-tag, Lex A DNA binding domain (DBD) fusions, GAL4 DNA binding domain fusions, and herpes simplex virus (HSV) BP 16 protein fusions. A fusion protein also can be engineered to contain a cleavage site located between the elongase HSEL01-like protein polypeptide-encoding sequence and the heterologous protein sequence, so that the elongase HSEL01-like protein polypeptide can be cleaved and purified away from the heterologous moiety.

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A fusion protein can be synthesized chemically, as is known in the art. Preferably, a fusion protein is produced by covalently linking two polypeptide segments or by standard procedures in the art of molecular biology. Recombinant DNA methods can be used to prepare fusion proteins, for example, by making a DNA construct which comprises coding sequences selected from SEQ ID NO: 1 in proper reading frame with nucleotides encoding the second polypeptide segment and expressing the DNA construct in a host cell, as is known in the art. Many kits for constructing fusion proteins are available from companies such as Promega Corporation (Madison, WI), Stratagene (La Jolla, CA), CLONTECH (Mountain View, CA), Santa Cruz Biotechnology (Santa Cruz, CA), MBL International Corporation (Wellesley, MA), and Quantum Biotechnologies (Montreal, Canada; 1-888-DNA-KITS).

Identification of Species Homologs

25 Species homologs of human elongase HSEL01-like protein polypeptide can be obtained using elongase HSEL01-like protein polypeptide polynucleotides (described below) to make suitable probes or primers for screening cDNA expression libraries from other species, such as mice, monkeys, or yeast, identifying cDNAs which encode homologs of elongase HSEL01-like protein polypeptide, and expressing the cDNAs as is known in the art.

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Polynucleotides

An elongase HSEL01-like protein polynucleotide can be single- or double-stranded and comprises a coding sequence or the complement of a coding sequence for an elongase HSEL01-like protein polypeptide. Degenerate nucleotide sequences encoding human elongase HSEL01-like protein polypeptides, as well as homologous nucleotide sequences which are at least about 50, 55, 60, 65, 70, preferably about 75, 90, 95, 98, or 99% identical to the nucleotide sequence shown in SEQ ID NO: 1, 5, or 6 or the complement thereof also are elongase HSEL01-like protein polynucleotides. Percent sequence identity between the sequences of two polynucleotides is determined using computer programs such as ALIGN which employ the FASTA algorithm, using an affine gap search with a gap open penalty of -12 and a gap extension penalty of -2. Complementary DNA (cDNA) molecules, species homologs, and variants of elongase HSEL01-like protein polynucleotides that encode biologically active elongase HSEL01-like protein polypeptides also are elongase HSEL01-like protein polynucleotides. Polynucleotide fragments comprising at least 8, 9, 10, 11, 12, 15, 20, or 25 contiguous nucleotides of SEQ ID NO: 1, 5, or 6 or the complement thereof also are elongase HSEL01-like protein polynucleotides. These fragments can be used, for example, as hybridization probes or as antisense oligonucleotides.

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25 Variants and homologs of the elongase HSEL01-like protein polynucleotides described above also are elongase HSEL01-like protein polynucleotides. Typically, homologous elongase HSEL01-like protein polynucleotide sequences can be identified by hybridization of candidate polynucleotides to known elongase HSEL01-like protein polynucleotides under stringent conditions, as is known in the art. For example, using the following wash conditions—2X SSC (0.3 M NaCl, 0.03 M sodium citrate, pH 7.0), 0.1% SDS, room temperature twice, 30 minutes each; then 2X SSC, 0.1% SDS, 50°C once, 30 minutes; then 2X SSC, room

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temperature twice, 10 minutes each—homologous sequences can be identified which contain at most about 25-30% basepair mismatches. More preferably, homologous nucleic acid strands contain 15-25% basepair mismatches, even more preferably 5-15% basepair mismatches.

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Species homologs of the elongase HSEL01-like protein polynucleotides disclosed herein also can be identified by making suitable probes or primers and screening cDNA expression libraries from other species, such as mice, monkeys, or yeast. Human variants of elongase HSEL01-like protein polynucleotides can be identified, for example, by screening human cDNA expression libraries. It is well known that the T_m of a double-stranded DNA decreases by 1-1.5°C with every 1% decrease in homology (Bonner *et al*, *J. Mol. Biol.* 81, 123 (1973). Variants of human elongase HSEL01-like protein polynucleotides or elongase HSEL01-like protein polynucleotides of other species can therefore be identified by hybridizing a putative homologous elongase HSEL01-like protein polynucleotide with a polynucleotide having a nucleotide sequence of SEQ ID NO: 1, 5, or 6 or the complement thereof to form a test hybrid. The melting temperature of the test hybrid is compared with the melting temperature of a hybrid comprising polynucleotides having perfectly complementary nucleotide sequences, and the number or percent of basepair mismatches within the test hybrid is calculated.

Identification of Polynucleotide Variants and Homologs

Nucleotide sequences which hybridize to elongase HSEL01-like protein polynucleotides or their complements following stringent hybridization and/or wash conditions also are elongase HSEL01-like protein polynucleotides. Stringent wash conditions are well known and understood in the art and are disclosed, for example, in Sambrook *et al*, MOLECULAR CLONING: A LABORATORY MANUAL, 2d ed., 1989, at pages 9.50-9.51.

Typically, for stringent hybridization conditions a combination of temperature and salt concentration should be chosen that is approximately 12-20°C below the calculated T_m of the hybrid under study. The T_m of a hybrid between an elongase

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HSEL01-like protein polynucleotide having a nucleotide sequence shown in SEQ ID NO: 1, 5, or 6 or the complement thereof and a polynucleotide sequence which is at least about 50, preferably about 75, 90, 96, or 98% identical to one of those nucleotide sequences can be calculated, for example, using the equation of Bolton and McCarthy, *Proc. Natl. Acad. Sci. U.S.A.* 48, 1390 (1962):

$$T_m = 81.5^\circ\text{C} \cdot 16.6(\log_{10}[\text{Na}^+]) + 0.41(\%G + C) - 0.63(\%\text{formamide}) - 600/l,$$

where l = the length of the hybrid in basepairs.

Stringent wash conditions include, for example, 4X SSC at 65°C, or 50% formamide, 4X SSC at 42°C, or 0.5X SSC, 0.1% SDS at 65°C. Highly stringent wash conditions include, for example, 0.2X SSC at 65 °C.

Preparation of Polynucleotides

5 An elongase HSEL01-like protein polynucleotide can be isolated free of other cellular components such as membrane components, proteins, and lipids. Polynucleotides can be made by a cell and isolated using standard nucleic acid purification techniques, or synthesized using an amplification technique, such as the polymerase chain reaction (PCR), or by using an automatic synthesizer. Methods for isolating polynucleotides are routine and are known in the art. Any such technique for obtaining a polynucleotide can be used to obtain isolated elongase HSEL01-like protein polynucleotides. For example, restriction enzymes and probes can be used to isolate polynucleotide fragments, which comprise elongase HSEL01-like protein nucleotide sequences. Isolated polynucleotides are in preparations that are free or at least 70, 80, or 90% free of other molecules.

10 Human elongase HSEL01-like protein cDNA molecules can be made with standard molecular biology techniques, using elongase HSEL01-like protein mRNA as a template. Human elongase HSEL01-like protein cDNA molecules can thereafter be replicated using molecular biology techniques known in the art and disclosed in

15 An elongase HSEL01-like protein polynucleotide can be isolated free of other cellular components such as membrane components, proteins, and lipids. Polynucleotides can be made by a cell and isolated using standard nucleic acid purification techniques, or synthesized using an amplification technique, such as the polymerase chain reaction (PCR), or by using an automatic synthesizer. Methods for isolating polynucleotides are routine and are known in the art. Any such technique for obtaining a polynucleotide can be used to obtain isolated elongase HSEL01-like protein polynucleotides. For example, restriction enzymes and probes can be used to isolate polynucleotide fragments, which comprise elongase HSEL01-like protein nucleotide sequences. Isolated polynucleotides are in preparations that are free or at least 70, 80, or 90% free of other molecules.

20 Human elongase HSEL01-like protein cDNA molecules can be made with standard molecular biology techniques, using elongase HSEL01-like protein mRNA as a template. Human elongase HSEL01-like protein cDNA molecules can thereafter be replicated using molecular biology techniques known in the art and disclosed in

25 Various PCR-based methods can be used to extend the nucleic acid sequences disclosed herein to detect upstream sequences such as promoters and regulatory elements. For example, restriction-site PCR uses universal primers to retrieve unknown sequence adjacent to a known locus (Sarkar, *PCR Methods Applic.*, 2, 318-322, 1993). Genomic DNA is first amplified in the presence of a primer to a linker sequence and a primer specific to the known region. The amplified sequences are then subjected to a second round of PCR with the same linker primer and another specific primer internal to the first one. Products of each round of PCR are transcribed with an appropriate RNA polymerase and sequenced using reverse transcriptase.

30 Inverse PCR also can be used to amplify or extend sequences using divergent primers based on a known region (Triglia et al., *Nucleic Acids Res.* 16, 8186, 1988). Primers can be designed using commercially available software, such as OLIGO 4.06 Primer Analysis software (National Biosciences Inc., Plymouth, Minn.), to be 22-30 nucleotides in length, to have a GC content of 50% or more, and to anneal to the target sequence at temperatures about 68-72°C. The method uses several restriction enzymes to generate a suitable fragment in the known region of a gene. The fragment is then circularized by intramolecular ligation and used as a PCR template.

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Another method which can be used is capture PCR, which involves PCR amplification of DNA fragments adjacent to a known sequence in human and yeast artificial chromosome DNA (Lagerstrom et al., *PCR Methods Applic.*, 1, 111-119, 1991). In this method, multiple restriction enzyme digestions and ligation also can be used to place an engineered double-stranded sequence into an unknown fragment of the DNA molecule before performing PCR.

Another method which can be used to retrieve unknown sequences is that of Parker et al., *Nucleic Acids Res.* 19, 3055-3060, 1991). Additionally, PCR, nested primers, and PROMOTERENDER libraries (CLONTECH, Palo Alto, Calif.) can be used to walk genomic DNA (CLONTECH, Palo Alto, Calif.). This process avoids the need to screen libraries and is useful in finding intron/exon junctions.

When screening for full-length cDNAs, it is preferable to use libraries that have been size-selected to include larger cDNAs. Randomly-primed libraries are preferable, in that they will contain more sequences which contain the 5' regions of genes. Use of a randomly primed library may be especially preferable for situations in which an oligo d(T) library does not yield a full-length cDNA. Genomic libraries can be useful for extension of sequence into 5' non-transcribed regulatory regions.

Commercially available capillary electrophoresis systems can be used to analyze the size or confirm the nucleotide sequence of PCR or sequencing products. For example, capillary sequencing can employ flowable polymers for electrophoretic separation, four different fluorescent dyes (one for each nucleotide) that are laser activated, and detection of the emitted wavelengths by a charge coupled device camera. Output/light intensity can be converted to electrical signal using appropriate software (e.g. GENOTYPER and Sequence NAVIGATOR, Perkin Elmer), and the entire process from loading of samples to computer analysis and electronic data display can be computer controlled. Capillary electrophoresis is especially preferable

for the sequencing of small pieces of DNA that might be present in limited amounts in a particular sample.

Obtaining Polypeptides

Human elongase HSEL01-like protein polypeptides can be obtained, for example, by purification from human cells, by expression of elongase HSEL01-like protein polymucleotides, or by direct chemical synthesis.

Protein Purification

Human elongase HSEL01-like protein polypeptides can be purified from any cell that expresses the polypeptide, including host cells that have been transfected with elongase HSEL01-like protein expression constructs. A purified elongase HSEL01-like protein polypeptide is separated from other compounds that normally associate with the elongase HSEL01-like protein polypeptide in the cell, such as certain proteins, carbohydrates, or lipids, using methods well-known in the art. Such methods include, but are not limited to, size exclusion chromatography, ammonium sulfate fractionation, ion exchange chromatography, affinity chromatography, and preparative gel electrophoresis. A preparation of purified elongase HSEL01-like protein polypeptides is at least 80% pure; preferably, the preparations are 90%, 95%, or 99% pure. Purity of the preparations can be assessed by any means known in the art, such as SDS-polyacrylamide gel electrophoresis.

Expression of Polynucleotides

To express an elongase HSEL01-like protein polynucleotide, the polynucleotide can be inserted into an expression vector that contains the necessary elements for the transcription and translation of the inserted coding sequence. Methods that are well known to those skilled in the art can be used to construct expression vectors containing sequences encoding elongase HSEL01-like protein polypeptides and

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appropriate transcriptional and translational control elements. These methods include *in vitro* recombinant DNA techniques, synthetic techniques, and *in vivo* genetic recombination. Such techniques are described, for example, in Sambrook *et al.* (1989) and in Ausubel *et al.*, CURRENT PROTOCOLS IN MOLECULAR BIOLOGY, John Wiley & Sons, New York, N.Y., 1989.

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A variety of expression vector/host systems can be utilized to contain and express sequences encoding an elongase HSEL01-like protein polypeptide. These include, but are not limited to, microorganisms, such as bacteria transformed with recombinant bacteriophage, plasmid, or cosmid DNA expression vectors; yeast transformed with yeast expression vectors, insect cell systems infected with virus expression vectors (e.g., baculovirus), plant cell systems transformed with virus expression vectors (e.g., cauliflower mosaic virus, CaMV; tobacco mosaic virus, TMV) or with bacterial expression vectors (e.g., Ti or pBR322 plasmids), or animal cell systems.

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The control elements or regulatory sequences are those non-translated regions of the vector -- enhancers, promoters, 5' and 3' untranslated regions -- which interact with host cellular proteins to carry out transcription and translation. Such elements can vary in their strength and specificity. Depending on the vector system and host utilized, any number of suitable transcription and translation elements, including constitutive and inducible promoters, can be used. For example, when cloning in bacterial systems, inducible promoters such as the hybrid lacZ promoter of the BLUESCRIPT phagemid (Stratagene, LaJolla, Calif.) or pSPORT1 plasmid (Life Technologies) and the like can be used. The baculovirus polyhedrin promoter can be used in insect cells. Promoters or enhancers derived from the genomes of plant cells (e.g., heat shock, RUBISCO, and storage protein genes) or from plant viruses (e.g., viral promoters or leader sequences) can be cloned into the vector. In mammalian cell systems, promoters from mammalian genes or from mammalian viruses are preferable. If it is necessary to generate a cell line that contains multiple copies of a

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nucleotide sequence encoding an elongase HSEL01-like protein polypeptide, vectors based on SV40 or EBV can be used with an appropriate selectable marker.

Bacterial and Yeast Expression Systems

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In bacterial systems, a number of expression vectors can be selected depending upon the use intended for the elongase HSEL01-like protein polypeptide. For example, when a large quantity of an elongase HSEL01-like protein polypeptide is needed for the induction of antibodies, vectors which direct high level expression of fusion proteins that are readily purified can be used. Such vectors include, but are not limited to, multifunctional *E. coli* cloning and expression vectors such as BLUESCRIPT (Stratagene). In a BLUESCRIPT vector, a sequence encoding the elongase HSEL01-like protein polypeptide can be ligated into the vector in frame with sequences for the amino-terminal Met and the subsequent 7 residues of β -galactosidase so that a hybrid protein is produced. pIN vectors (Van Heeke & Schuster, *J. Biol. Chem.* 264, 5503-5509, 1989) or pGEX vectors (Prongea, Madison, Wis.) also can be used to express foreign polypeptides as fusion proteins with glutathione S-transferase (GST). In general, such fusion proteins are soluble and can easily be purified from lysed cells by adsorption to glutathione-agarose beads followed by elution in the presence of free glutathione. Proteins made in such systems can be designed to include heparin, thrombin, or factor Xa protease cleavage sites so that the cloned polypeptide of interest can be released from the GST moiety at will.

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In the yeast *Saccharomyces cerevisiae*, a number of vectors containing constitutive or inducible promoters such as alpha factor, alcohol oxidase, and PGH can be used. For reviews, see Ausubel *et al.* (1989) and Grant *et al.*, *Methods Enzymol.* 153, 516-544, 1987.

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Plant and Insect Expression Systems

If plant expression vectors are used, the expression of sequences encoding elongase HSELO1-like protein polypeptides can be driven by any of a number of promoters. For example, viral promoters such as the 3S and 19S promoters of CaMV can be used alone or in combination with the omega leader sequence from TMV (Takamatsu, *EMBO J.* 6, 307-311, 1987). Alternatively, plant promoters such as the small subunit of RUBISCO or heat shock promoters can be used (Coruzzi *et al.*, *EMBO J.* 3, 1671-1680, 1984; Broglie *et al.*, *Science* 224, 838-843, 1984; Winter *et al.*, *Results Probl. Cell Differ.* 17, 85-105, 1991). These constructs can be introduced into plant cells by direct DNA transformation or by pathogen-mediated transfection. Such techniques are described in a number of generally available reviews (e.g., Hobbs or Murray, in *McGRAW HILL YEARBOOK OF SCIENCE AND TECHNOLOGY*, McGraw Hill, New York, N.Y., pp. 191-196, 1992).

An insect system also can be used to express an elongase HSELO1-like protein polypeptide. For example, in one such system *Autographa californica* nuclear polyhedrosis virus (AcNPV) is used as a vector to express foreign genes in *Spodoptera frugiperda* cells or in *Trichoplusia* larvae. Sequences encoding elongase HSELO1-like protein polypeptides can be cloned into a non-essential region of the virus, such as the polyhedrin gene, and placed under control of the polyhedrin promoter. Successful insertion of elongase HSELO1-like protein polypeptides will render the polyhedrin gene inactive and produce recombinant virus lacking coat protein. The recombinant viruses can then be used to infect *S. frugiperda* cells or *Trichoplusia* larvae in which elongase HSELO1-like protein polypeptides can be expressed (Engelhard *et al.*, *Proc. Natl. Acad. Sci.* 91, 3224-3227, 1994).

Mammalian Expression Systems

A number of viral-based expression systems can be used to express elongase HSELO1-like protein polypeptides in mammalian host cells. For example, if an

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adenovirus is used as an expression vector, sequences encoding elongase HSELO1-like protein polypeptides can be ligated into an adenovirus transcription/translation complex comprising the late promoter and tripartite leader sequence. Insertion in a non-essential E1 or E3 region of the viral genome can be used to obtain a viable virus that is capable of expressing an elongase HSELO1-like protein polypeptide in infected host cells (Logan & Shenk, *Proc. Natl. Acad. Sci.* 81, 3655-3659, 1984). If desired, transcription enhancers, such as the Rous sarcoma virus (RSV) enhancer, can be used to increase expression in mammalian host cells.

5 Human artificial chromosomes (HACs) also can be used to deliver larger fragments of DNA than can be contained and expressed in a plasmid. HACs of 6M to 10M are constructed and delivered to cells via conventional delivery methods (e.g., liposomes, polycationic amino polymers, or vesicles).

10 Specific initiation signals also can be used to achieve more efficient translation of sequences encoding elongase HSELO1-like protein polypeptides. Such signals include the ATG initiation codon and adjacent sequences. In cases where sequences encoding an elongase HSELO1-like protein polypeptide, its initiation codon, and upstream sequences are inserted into the appropriate expression vector, no additional transcriptional or translational control signals may be needed. However, in cases where only coding sequence, or a fragment thereof, is inserted, exogenous translational control signals (including the ATG initiation codon) should be provided. The initiation codon should be in the correct reading frame to ensure translation of the entire insert. Exogenous translational elements and initiation codons can be of various origins, both natural and synthetic. The efficiency of expression can be enhanced by the inclusion of enhancers which are appropriate for the particular cell system which is used (see Scharf *et al.*, *Results Probl. Cell Differ.* 20, 125-162, 1994).

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Host Cells

A host cell strain can be chosen for its ability to modulate the expression of the inserted sequences or to process the expressed elongase HSEL01-like protein polypeptide in the desired fashion. Such modifications of the polypeptide include, but are not limited to, acetylation, carboxylation, glycosylation, phosphorylation, lipidation, and acylation. Post-translational processing which cleaves a "prepro" form of the polypeptide also can be used to facilitate correct insertion, folding and/or function. Different host cells that have specific cellular machinery and characteristic mechanisms for post-translational activities (e.g., CHO, HeLa, MDCK, HEK293, and WI38), are available from the American Type Culture Collection (ATCC; 10801 University Boulevard, Manassas, VA 20110-2209) and can be chosen to ensure the correct modification and processing of the foreign protein.

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Stable expression is preferred for long-term, high-yield production of recombinant proteins. For example, cell lines which stably express elongase HSEL01-like protein polypeptides can be transformed using expression vectors which can contain viral origins of replication and/or endogenous expression elements and a selectable marker gene on the same or on a separate vector. Following the introduction of the vector, cells can be allowed to grow for 1-2 days in an enriched medium before they are switched to a selective medium. The purpose of the selectable marker is to confer resistance to selection, and its presence allows growth and recovery of cells which successfully express the introduced elongase HSEL01-like protein sequences. Resistant clones of stably transformed cells can be proliferated using tissue culture techniques appropriate to the cell type. See, for example, ANIMAL CELL CULTURE, R.I. Freshney, ed., 1986.

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Detecting Expression

Although the presence of marker gene expression suggests that the elongase HSEL01-like protein polynucleotide is also present, its presence and expression may need to be confirmed. For example, if a sequence encoding an elongase HSEL01-like protein polypeptide is inserted within a marker gene sequence, transformed cells containing sequences that encode an elongase HSEL01-like protein polypeptide can be identified by the absence of marker gene function. Alternatively, a marker gene can be placed in tandem with a sequence encoding an elongase HSEL01-like protein polypeptide under the control of a single promoter. Expression of the marker gene in response to induction or selection usually indicates expression of the elongase HSEL01-like protein polynucleotide.

Any number of selection systems can be used to recover transformed cell lines.

These include, but are not limited to, the herpes simplex virus thymidine kinase (Wigler et al., *Cell* 11, 223-32, 1977) and adenine phosphoribosyltransferase (Lowy

et al., *Cell* 22, 817-23, 1980) genes which can be employed in *tk* or *aprl* cells, respectively. Also, antimetabolite, antibiotic, or herbicide resistance can be used as the basis for selection. For example, *dfr5* confers resistance to methotrexate (Wigler et al., *Proc. Natl. Acad. Sci.* 77, 3567-70, 1980), *npl* confers resistance to the aminoglycosides, neomycin and G-418 (Colbere-Garapin et al., *J. Mol. Biol.* 150, 1-14, 1981), and *als* and *par* confer resistance to chlorsulfuron and phosphinotricin acetyltransferase, respectively (Murray, 1992, *supra*). Additional selectable genes have been described. For example, *trpB* allows cells to utilize indole in place of tryptophan, or *hisD*, which allows cells to utilize histidol in place of histidine (Hartman & Mulligan, *Proc. Natl. Acad. Sci.* 85, 8047-51, 1988). Visible markers such as anthocyanins, β -glucuronidase and its substrate GUS, and luciferase and its substrate luciferin, can be used to identify transformants and to quantify the amount of transient or stable protein expression attributable to a specific vector system (Rhodes et al., *Methods Mol. Biol.* 55, 121-131, 1995).

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procedures include, but are not limited to, DNA-DNA or DNA-RNA hybridizations and protein bioassay or immunoassay techniques that include membrane, solution, or chip-based technologies for the detection and/or quantification of nucleic acid or protein. For example, the presence of a polynucleotide sequence encoding an elongase HSELO1-like protein polypeptide can be detected by DNA-DNA or DNA-RNA hybridization or amplification using probes or fragments or fragments of polynucleotides encoding an elongase HSELO1-like protein polypeptide. Nucleic acid amplification-based assays involve the use of oligonucleotides selected from sequences encoding an elongase HSELO1-like protein polypeptide to detect transformants that contain an elongase HSELO1-like protein polynucleotide.

A variety of protocols for detecting and measuring the expression of an elongase HSELO1-like protein polypeptide, using either polyclonal or monoclonal antibodies specific for the polypeptide, are known in the art. Examples include enzyme-linked immunosorbent assay (ELISA), radioimmunoassay (RIA), and fluorescence activated cell sorting (FACS). A two-site, monoclonal-based immunoassay using monoclonal antibodies reactive to two non-interfering epitopes on an elongase HSELO1-like protein polypeptide can be used, or a competitive binding assay can be employed. These and other assays are described in Hampton *et al.*, SEROLOGICAL METHODS: A LABORATORY MANUAL, APS Press, St. Paul, Minn., 1990) and Maddox *et al.*, *J. Exp. Med.* 158, 1211-1216, 1983).

A wide variety of labels and conjugation techniques are known by those skilled in the art and can be used in various nucleic acid and amino acid assays. Means for producing labeled hybridization or PCR probes for detecting sequences related to polynucleotides encoding elongase HSELO1-like protein polypeptides include oligolabeling, nick translation, end-labeling, or PCR amplification using a labeled nucleotide. Alternatively, sequences encoding an elongase HSELO1-like protein polypeptide can be cloned into a vector for the production of an mRNA probe. Such vectors are known in the art, are commercially available, and can be used to synthesize RNA probes *in vitro* by addition of labeled nucleotides and an appropriate

RNA polymerase such as T7, T3, or SP6. These procedures can be conducted using a variety of commercially available kits (Amersham Pharmacia Biotech, Promega, and US Biochemical). Suitable reporter molecules or labels which can be used for ease of detection include radionuclides, enzymes, and fluorescent, chemiluminescent, or chromogenic agents, as well as substrates, cofactors, inhibitors, magnetic particles, and the like.

Expression and Purification of Polypeptides

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Host cells transformed with nucleotide sequences encoding an elongase HSELO1-like protein polypeptide can be cultured under conditions suitable for the expression and recovery of the protein from cell culture. The polypeptide produced by a transformed cell can be secreted or contained intracellularly depending on the sequence and/or the vector used. As will be understood by those of skill in the art, expression vectors containing poly nucleotides which encode elongase HSELO1-like protein polypeptides can be designed to contain signal sequences which direct secretion of soluble elongase HSELO1-like protein polypeptides through a prokaryotic or eukaryotic cell membrane or which direct the membrane insertion of membrane-bound elongase HSELO1-like protein polypeptide.

As discussed above, other constructions can be used to join a sequence encoding an elongase HSELO1-like protein polypeptide to a nucleotide sequence encoding a polypeptide domain which will facilitate purification of soluble proteins. Such purification facilitating domains include, but are not limited to, metal chelating peptides such as histidine-tryptophan modules that allow purification on immobilized metals, protein A domains that allow purification on immobilized immunoglobulin, and the domain utilized in the FLAG extension/affinity purification system (Immunex Corp., Seattle, Wash.). Inclusion of cleavable linker sequences such as those specific for Factor Xa or enterokinase (Invitrogen, San Diego, CA) between the purification domain and the elongase HSELO1-like protein polypeptide also can be used to facilitate purification. One such expression vector provides for expression of

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a fusion protein containing an elongase HSEL01-like protein polypeptide and 6 histidine residues preceding a thiodoxin or an enterokinase cleavage site. The histidine residues facilitate purification by IMAC (immobilized metal ion affinity chromatography, as described in Porath *et al.*, *Prot. Exp. Purif.* 3, 263-281, 1992), while the enterokinase cleavage site provides a means for purifying the elongase HSEL01-like protein polypeptide from the fusion protein. Vectors that contain fusion proteins are disclosed in Kroll *et al.*, *DNA Cell Biol.* 12, 441-453, 1993.

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Chemical Synthesis

Sequences encoding an elongase HSEL01-like protein polypeptide can be synthesized, in whole or in part, using chemical methods well known in the art (see Carruthers *et al.*, *Nucl. Acids Res. Symp. Ser.* 215-223, 1980; Horn *et al.*, *Nucl. Acids Res. Symp. Ser.* 225-232, 1980). Alternatively, an elongase HSEL01-like protein polypeptide itself can be produced using chemical methods to synthesize its amino acid sequence, such as by direct peptide synthesis using solid-phase techniques (Merifield, *J. Am. Chem. Soc.* 85, 2149-2154, 1963; Roberge *et al.*, *Science* 269, 202-204, 1995). Protein synthesis can be performed using manual techniques or by automation. Automated synthesis can be achieved, for example, using Applied Biosystems 431A Peptide Synthesizer (Perkin Elmer). Optionally, fragments of elongase HSEL01-like protein polypeptides can be separately synthesized and combined using chemical methods to produce a full-length molecule.

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The newly synthesized peptide can be substantially purified by preparative high performance liquid chromatography (e.g., Creighton, *PROTEINS: STRUCTURES AND MOLECULAR PRINCIPLES*, WH Freeman and Co., New York, N.Y., 1983). The composition of a synthetic elongase HSEL01-like protein polypeptide can be confirmed by amino acid analysis or sequencing (e.g., the Edman degradation procedure; see Creighton, *supra*). Additionally, any portion of the amino acid sequence of the elongase HSEL01-like protein polypeptide can be altered during

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25 performance liquid chromatography (e.g., Creighton, *PROTEINS: STRUCTURES AND MOLECULAR PRINCIPLES*, WH Freeman and Co., New York, N.Y., 1983). The composition of a synthetic elongase HSEL01-like protein polypeptide can be confirmed by amino acid analysis or sequencing (e.g., the Edman degradation procedure; see Creighton, *supra*). Additionally, any portion of the amino acid sequence of the elongase HSEL01-like protein polypeptide can be altered during

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direct synthesis and/or combined using chemical methods with sequences from other proteins to produce a variant polypeptide or a fusion protein.

Production of Altered Polypeptides

As will be understood by those of skill in the art, it may be advantageous to produce elongase HSEL01-like protein polypeptide-encoding nucleotide sequences possessing non-naturally occurring codons. For example, codons preferred by a particular prokaryotic or eukaryotic host can be selected to increase the rate of protein expression or to produce an RNA transcript having desirable properties, such as a half-life that is longer than that of a transcript generated from the naturally occurring sequence.

The nucleotide sequences disclosed herein can be engineered using methods generally known in the art to alter elongase HSEL01-like protein polypeptide-encoding sequences for a variety of reasons, including but not limited to, alterations which modify the cloning, processing, and/or expression of the polypeptide or mRNA product. DNA shuffling by random fragmentation and PCR reassembly of gene fragments and synthetic oligonucleotides can be used to engineer the nucleotide sequences. For example, site-directed mutagenesis can be used to insert new restriction sites, alter glycosylation patterns, change codon preference, produce splice variants, introduce mutations, and so forth.

Antibodies

Any type of antibody known in the art can be generated to bind specifically to an epitope of an elongase HSEL01-like protein polypeptide. "Antibody" as used herein includes intact immunoglobulin molecules, as well as fragments thereof, such as Fab, F(ab')₂, and Fv, which are capable of binding an epitope of an elongase HSEL01-like protein polypeptide. Typically, at least 6, 8, 10, or 12 contiguous amino acids

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are required to form an epitope. However, epitopes which involve non-contiguous amino acids may require more, e.g., at least 15, 25, or 50 amino acids.

An antibody which specifically binds to an epitope of an elongase HSEL01-like protein polypeptide can be used therapeutically, as well as in immunochemical assays, such as Western blots, ELISAs, radioimmunoassays, immunohistochemical assays, immunoprecipitations, or other immunochemical assays known in the art. Various immunoassays can be used to identify antibodies having the desired specificity. Numerous protocols for competitive binding or immunoradiometric assays are well known in the art. Such immunoassays typically involve the measurement of complex formation between an immunogen and an antibody that specifically binds to the immunogen.

Typically, an antibody which specifically binds to an elongase HSEL01-like protein polypeptide provides a detection signal at least 5-, 10-, or 20-fold higher than a detection signal provided with other proteins when used in an immunochemical assay. Preferably, antibodies which specifically bind to elongase HSEL01-like protein polypeptides do not detect other proteins in immunochemical assays and can immunoprecipitate an elongase HSEL01-like protein polypeptide from solution.

Human elongase HSEL01-like protein polypeptides can be used to immunize a mammal, such as a mouse, rat, rabbit, guinea pig, monkey, or human, to produce polyclonal antibodies. If desired, an elongase HSEL01-like protein polypeptide can be conjugated to a carrier protein, such as bovine serum albumin, thyroglobulin, and keyhole limpet hemocyanin. Depending on the host species, various adjuvants can be used to increase the immunological response. Such adjuvants include, but are not limited to, Freund's adjuvant, mineral gels (e.g., aluminum hydroxide), and surface active substances (e.g., lysolecithin, pluronic polyols, polyanions, peptides, oil emulsions, keyhole limpet hemocyanin, and dinitrophenol). Among adjuvants used in humans, BCG (*bacilli Calmette-Guerin*) and *Corynebacterium parvum* are especially useful.

Monoclonal antibodies that specifically bind to an elongase HSEL01-like protein polypeptide can be prepared using any technique which provides for the production of antibody molecules by continuous cell lines in culture. These techniques include, but are not limited to, the hybridoma technique, the human B-cell hybridoma technique, and the EBV-hybridoma technique (Kohler *et al.*, *Nature* 256, 495-497, 1985; Kozbor *et al.*, *J. Immunol. Methods* 81, 31-42, 1985; Cote *et al.*, *Proc. Natl. Acad. Sci. USA* 80, 2026-2030, 1983; Cole *et al.*, *Mol. Cell Biol.* 62, 109-120, 1984).

In addition, techniques developed for the production of "chimeric antibodies," the splicing of mouse antibody genes to human antibody genes to obtain a molecule with appropriate antigen specificity and biological activity, can be used (Morrison *et al.*, *Proc. Natl. Acad. Sci. USA* 81, 6831-6835, 1984; Neuberger *et al.*, *Nature* 312, 604-608, 1984; Takeda *et al.*, *Nature* 314, 452-454, 1985). Monoclonal and other antibodies also can be "humanized" to prevent a patient from mounting an immune response against the antibody when it is used therapeutically. Such antibodies may be sufficiently similar in sequence to human antibodies to be used directly in therapy or may require alteration of a few key residues. Sequence differences between rodent antibodies and human sequences can be minimized by replacing residues which differ from those in the human sequences by site directed mutagenesis of individual residues or by grafting of entire complementarity determining regions. Alternatively, humanized antibodies can be produced using recombinant methods, as described in GB2183638B. Antibodies that specifically bind to an elongase HSEL01-like protein polypeptide can contain antigen binding sites which are either partially or fully humanized, as disclosed in U.S. 5,565,332.

Alternatively, techniques described for the production of single chain antibodies can be adapted using methods known in the art to produce single chain antibodies that specifically bind to elongase HSEL01-like protein polypeptides. Antibodies with related specificity, but of distinct idiotypic composition, can be generated by chain

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shuffling from random combinatorial immunoglobulin libraries (Burton, *Proc. Natl. Acad. Sci.* **88**, 11120-23, 1991).

Single-chain antibodies also can be constructed using a DNA amplification method, such as PCR, using hybridoma cDNA as a template (Thirion *et al.*, 1996, *Eur. J. Cancer Prev.* **5**, S07-11). Single-chain antibodies can be mono- or bispecific, and can be bivalent or tetravalent. Construction of trivalent, bispecific single-chain antibodies is taught, for example, in Coloma & Morrison, 1997, *Nat. Biotechnol.* **15**, 159-63. Construction of bivalent, bispecific single-chain antibodies is taught in Mallender & Voss, 1994, *J. Biol. Chem.* **269**, 199-206.

A nucleotide sequence encoding a single-chain antibody can be constructed using manual or automated nucleotide synthesis, cloned into an expression construct using standard recombinant DNA methods, and introduced into a cell to express the coding sequence, as described below. Alternatively, single-chain antibodies can be produced directly using, for example, filamentous phage technology (Verhaar *et al.*, 1995, *Int. J. Cancer* **61**, 497-501; Nicholls *et al.*, 1993, *J. Immunol. Meth.* **165**, 81-91).

Antibodies which specifically bind to elongase HSEL01-like protein polypeptides also can be produced by inducing *in vivo* production in the lymphocyte population or by screening immunoglobulin libraries or panels of highly specific binding reagents as disclosed in the literature (Orlandi *et al.*, *Proc. Natl. Acad. Sci.* **86**, 3833-3837, 1989; Winter *et al.*, *Nature* **349**, 293-299, 1991).

Other types of antibodies can be constructed and used therapeutically in methods of the invention. For example, chimeric antibodies can be constructed as disclosed in WO 93/03151. Binding proteins which are derived from immunoglobulins and which are multivalent and multispecific, such as the "diabodies" described in WO 94/13804, also can be prepared.

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Antibodies according to the invention can be purified by methods well known in the art. For example, antibodies can be affinity purified by passage over a column to which an elongase HSEL01-like protein polypeptide is bound. The bound antibodies can then be eluted from the column using a buffer with a high salt concentration.

5 Antisense Oligonucleotides

Antisense oligonucleotides are nucleotide sequences that are complementary to a specific DNA or RNA sequence. Once introduced into a cell, the complementary nucleotides combine with natural sequences produced by the cell to form complexes and block either transcription or translation. Preferably, an antisense oligonucleotide is at least 11 nucleotides in length, but can be at least 12, 15, 20, 25, 30, 35, 40, 45, or 50 or more nucleotides long. Longer sequences also can be used. Antisense oligonucleotide molecules can be provided in a DNA construct and introduced into a cell as described above to decrease the level of elongase HSEL01-like protein gene products in the cell.

Antisense oligonucleotides can be deoxyribonucleotides, ribonucleotides, or a combination of both. Oligonucleotides can be synthesized manually or by an automated synthesizer, by covalently linking the 5' end of one nucleotide with the 3' end of another nucleotide with non-phosphodiester internucleotide linkages such as alkylphosphonates, phosphorothioates, phosphordithioates, alkylphosphonothioates, alkylphosphonates, phosphoramidates, phosphate esters, carbamates, acetamide, carboxymethyl esters, carbonates, and phosphate triesters. See Brown, *Meth. Mol. Biol.* **20**, 1-8, 1994; Sonveaux, *Meth. Mol. Biol.* **26**, 1-72, 1994; Uhmann *et al.*, *Chem. Rev.* **90**, 543-583, 1990.

Modifications of elongase HSEL01-like protein gene expression can be obtained by designing antisense oligonucleotides that will form duplexes to the control, 5', or regulatory regions of the elongase HSEL01-like protein gene. Oligonucleotides derived from the transcription initiation site, e.g., between positions -10 and +10

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from the start site, are preferred. Similarly, inhibition can be achieved using "triple helix" base-pairing methodology. Triple helix pairing is useful because it causes inhibition of the double helix to open sufficiently for the binding of polymerases, transcription factors, or chaperons. Therapeutic advances using triple DNA have been described in the literature (e.g., Gee *et al.*, in Huber & Carr, MOLECULAR AND IMMUNOLOGIC APPROACHES, Futura Publishing Co., Mt. Kisco, N.Y., 1994). An antisense oligonucleotide also can be designed to block translation of mRNA by preventing the transcript from binding to ribosomes.

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Ribozymes

phosphate group are substituted, also can be employed in a modified antisense oligonucleotide. These modified oligonucleotides can be prepared by methods well known in the art. See, e.g., Agrawal *et al.*, *Trends Biotechnol.* 10, 152-158, 1992; Uhlmann *et al.*, *Chem. Rev.* 90, 543-584, 1990; Uhlmann *et al.*, *Tetrahedron Lett.* 215, 3539-3542, 1987.

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Ribozymes are RNA molecules with catalytic activity. See, e.g., Cech, *Science* 236, 1532-1539; 1987; Cech, *Ann. Rev. Biochem.* 59, 543-568; 1990, Czech, *Curr. Opin. Struct. Biol.* 2, 605-609; 1992, Couture & Stinchcomb, *Trends Genet.* 12, 510-515, 1996. Ribozymes can be used to inhibit gene function by cleaving an RNA sequence, as is known in the art (e.g., Haseloff *et al.*, U.S. Patent 5,641,673). The mechanism of ribozyme action involves sequence-specific hybridization of the ribozyme molecule to complementary target RNA, followed by endonucleolytic cleavage. Examples include engineered hammerhead motif ribozyme molecules that can specifically and efficiently catalyze endonucleolytic cleavage of specific nucleotide sequences.

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Precise complementarity is not required for successful complex formation between an antisense oligonucleotide and the complementary sequence of an elongase HSEL01-like protein polynucleotide. Antisense oligonucleotides which comprise, for example, 2, 3, 4, or 5 or more stretches of contiguous nucleotides which are precisely complementary to an elongase HSEL01-like protein polynucleotide, each separated by a stretch of contiguous nucleotides which are not complementary to adjacent elongase HSEL01-like protein nucleotides, can provide sufficient targeting specificity for elongase HSEL01-like protein mRNA. Preferably, each stretch of complementary contiguous nucleotides is at least 4, 5, 6, 7, or 8 or more nucleotides in length. Non-complementary intervening sequences are preferably 1, 2, 3, or 4 nucleotides in length. One skilled in the art can easily use the calculated melting point of an antisense-sense pair to determine the degree of mismatching which will be tolerated between a particular antisense oligonucleotide and a particular elongase HSEL01-like protein polynucleotide sequence.

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Antisense oligonucleotides can be modified without affecting their ability to hybridize to an elongase HSEL01-like protein polynucleotide. These modifications can be internal or at one or both ends of the antisense molecule. For example, internucleoside phosphate linkages can be modified by adding cholesterol or diamine moieties with varying numbers of carbon residues between the amino groups and terminal ribose. Modified bases and/or sugars, such as arabinose instead of ribose, or a 3', 5'-substituted oligonucleotide in which the 3' hydroxyl group or the 5'

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The coding sequence of an elongase HSEL01-like protein polynucleotide can be used to generate ribozymes that will specifically bind to mRNA transcribed from the elongase HSEL01-like protein polynucleotide. Methods of designing and constructing ribozymes which can cleave other RNA molecules in trans in a highly sequence specific manner have been developed and described in the art (see Haseloff *et al.* *Nature* 334, 585-591, 1988). For example, the cleavage activity of ribozymes can be targeted to specific RNAs by engineering a discrete "hybridization" region into the ribozyme. The hybridization region contains a sequence complementary to the target RNA and thus specifically hybridizes with the target (see, for example, Gerlach *et al.*, EP 321,201).

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Antisense oligonucleotides can be modified without affecting their ability to hybridize to an elongase HSEL01-like protein polynucleotide. These modifications can be internal or at one or both ends of the antisense molecule. For example, internucleoside phosphate linkages can be modified by adding cholesterol or diamine moieties with varying numbers of carbon residues between the amino groups and terminal ribose. Modified bases and/or sugars, such as arabinose instead of ribose, or a 3', 5'-substituted oligonucleotide in which the 3' hydroxyl group or the 5'

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Specific ribozyme cleavage sites within an elongase HSELO1-like protein RNA target can be identified by scanning the target molecule for ribozyme cleavage sites which include the following sequences: GUA, GUU, and GUC. Once identified, short RNA sequences of between 15 and 20 ribonucleotides corresponding to the 5 region of the target RNA containing the cleavage site can be evaluated for secondary structural features which may render the target inoperable. Suitability of candidate elongase HSELO1-like protein RNA targets also can be evaluated by testing accessibility to hybridization with complementary oligonucleotides using ribonuclease protection assays. Longer complementary sequences can be used to increase the 10 affinity of the hybridization sequence for the target. The hybridizing and cleavage regions of the ribozyme can be integrally related such that upon hybridizing to the target RNA through the complementary regions, the catalytic region of the ribozyme can cleave the target.

15 Ribozymes can be introduced into cells as part of a DNA construct. Mechanical methods, such as microinjection, liposome-mediated transfection, electroporation, or calcium phosphate precipitation, can be used to introduce a ribozyme-containing DNA construct into cells in which it is desired to decrease elongase HSELO1-like protein expression. Alternatively, if it is desired that the cells stably retain the DNA construct, the construct can be supplied on a plasmid and maintained as a separate element or integrated into the genome of the cells, as is known in the art. A ribozyme-encoding DNA construct can include transcriptional regulatory elements, such as a promoter element, an enhancer or UAS element, and a transcriptional terminator signal, for controlling transcription of ribozymes in the cells.

20 As taught in Haseloff *et al.*, U.S. Patent 5,641,673, ribozymes can be engineered so that ribozyme expression will occur in response to factors that induce expression of a target gene. Ribozymes also can be engineered to provide an additional level of regulation, so that destruction of mRNA occurs only when both a ribozyme and a target gene are induced in the cells.

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Differentially Expressed Genes

Described herein are methods for the identification of genes whose products interact with human elongase HSELO1-like protein. Such genes may represent genes that are differentially expressed in disorders including, but not limited to, cancer, diabetes, and CNS disorders. Further, such genes may represent genes that are differentially regulated in response to manipulations relevant to the progression or treatment of such diseases. Additionally, such genes may have a temporally modulated expression, increased or decreased at different stages of tissue or organism development. A differentially expressed gene may also have its expression modulated under control versus experimental conditions. In addition, the human elongase HSELO1-like protein gene or gene product may itself be tested for differential expression.

The degree to which expression differs in a normal versus a diseased state need only 5 be large enough to be visualized via standard characterization techniques such as differential display techniques. Other such standard characterization techniques by which expression differences may be visualized include but are not limited to, quantitative RT (reverse transcriptase), PCR, and Northern analysis.

Identification of Differentially Expressed Genes

To identify differentially expressed genes total RNA or, preferably, mRNA is isolated from tissues of interest. For example, RNA samples are obtained from tissues of experimental subjects and from corresponding tissues of control subjects. Any 10 RNA isolation technique that does not select against the isolation of mRNA may be utilized for the purification of such RNA samples. See, for example, Ausubel *et al.*, ed., CURRENT PROTOCOLS IN MOLECULAR BIOLOGY, John Wiley & Sons, Inc. New York, 1987-1993. Large numbers of tissue samples may readily be processed using techniques well known to those of skill in the art, such as, for example, the single-step RNA isolation process of Chomczynski, U.S. Patent 4,843,155.

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Transcripts within the collected RNA samples that represent RNA produced by differentially expressed genes are identified by methods well known to those of skill in the art. They include, for example, differential screening (Tedder *et al.*, *Proc. Natl. Acad. Sci. U.S.A.* 85, 208-12, 1988), subtractive hybridization (Hedrick *et al.*, *Nature* 308, 149-53; Lee *et al.*, *Proc. Natl. Acad. Sci. U.S.A.* 88, 2825, 1984), and, preferably, differential display (Liang & Pardue, *Science* 237, 967-71, 1992; U.S. Patent 5,262,311).

The differential expression information may itself suggest relevant methods for the treatment of disorders involving the human elongase HSEL01-like protein. For example, treatment may include a modulation of expression of the differentially expressed genes and/or the gene encoding the human elongase HSEL01-like protein. The differential expression information may indicate whether the expression or activity of the differentially expressed gene or gene product or the human elongase HSEL01-like protein gene or gene product are up-regulated or down-regulated.

Screening Methods

The invention provides assays for screening test compounds that bind to or modulate the activity of an elongase HSEL01-like protein polypeptide or an elongase HSEL01-like protein polynucleotide. A test compound preferably binds to an elongase HSEL01-like protein polypeptide or polynucleotide. More preferably, a test compound decreases or increases enzyme activity by at least about 10, preferably about 50, more preferably about 75, 90, or 100% relative to the absence of the test compound.

Test Compounds

Test compounds can be pharmacologic agents already known in the art or can be compounds previously unknown to have any pharmacological activity. The compounds can be naturally occurring or designed in the laboratory. They can be

isolated from microorganisms, animals, or plants, and can be produced recombinantly, or synthesized by chemical methods known in the art. If desired, test compounds can be obtained using any of the numerous combinatorial library methods known in the art, including but not limited to, biological libraries, spatially addressable parallel solid phase or solution phase libraries, synthetic library methods requiring deconvolution, the "one-bead one-compound" library method, and synthetic library methods using affinity chromatography selection. The biological library approach is limited to polypeptide libraries, while the other four approaches are applicable to polypeptide, non-peptide oligomer, or small molecule libraries of compounds. See Lam, *Anticancer Drug Des.* 12, 145, 1997.

Methods for the synthesis of molecular libraries are well known in the art (see, for example, DeVitt *et al.*, *Proc. Natl. Acad. Sci. U.S.A.* 90, 6909, 1993; Brb *et al.*, *Proc. Natl. Acad. Sci. U.S.A.* 91, 11422, 1994; Zuckermann *et al.*, *J. Med. Chem.* 37, 2678, 1994; Cho *et al.*, *Science* 261, 1303, 1993; Carell *et al.*, *Angew. Chem. Int. Ed. Engl.* 33, 2059, 1994; Carell *et al.*, *Angew. Chem. Int. Ed. Engl.* 33, 2061; Gallop *et al.*, *J. Med. Chem.* 37, 1233, 1994). Libraries of compounds can be presented in solution (see, e.g., Houghten, *BioTechniques* 13, 412-421, 1992), or on beads (Lam, *Nature* 354, 82-84, 1991), chips (Fodor, *Nature* 364, 555-556, 1993), bacteria or spores (Ladner, U.S. Patent 5,223,409), plasmids (Cull *et al.*, *Proc. Natl. Acad. Sci. U.S.A.* 89, 1855-1869, 1992), or phage (Scott & Smith, *Science* 249, 386-390, 1990; Devin, *Science* 249, 404-406, 1990); Cwirla *et al.*, *Proc. Natl. Acad. Sci. 97*, 6378-6382, 1990; Felici, *J. Mol. Biol.* 222, 301-310, 1991; and Ladner, U.S. Patent 5,223,409).

High Throughput Screening

Test compounds can be screened for the ability to bind to elongase HSEL01-like protein polypeptides or polynucleotides or to affect elongase HSEL01-like protein activity or elongase HSEL01-like protein gene expression using high throughput screening. Using high throughput screening, many discrete compounds can be tested in parallel so that large numbers of test compounds can be quickly screened. The

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most widely established techniques utilize 96-well microtiter plates. The wells of the microtiter plates typically require assay volumes that range from 50 to 500 μ l. In addition to the plates, many instruments, materials, pipettors, robotics, plate washers, and plate readers are commercially available to fit the 96-well format.

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Alternatively, "free format assays," or assays that have no physical barrier between samples, can be used. For example, an assay using pigment cells (melanocytes) in a simple homogeneous assay for combinatorial peptide libraries is described by Jayawickreme *et al.*, *Proc. Natl. Acad. Sci. U.S.A.* 91, 1614-18 (1994). The cells are placed under agarose in petri dishes, then beads that carry combinatorial compounds are placed on the surface of the agarose. The combinatorial compounds are partially released the compounds from the beads. Active compounds can be visualized as dark pigment areas because, as the compounds diffuse locally into the gel matrix, the active compounds cause the cells to change colors.

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Another example of a free format assay is described by Chelsky, "Strategies for Screening Combinatorial Libraries: Novel and Traditional Approaches," reported at the First Annual Conference of The Society for Biomolecular Screening in Philadelphia, Pa. (Nov. 7-10, 1995). Chelsky placed a simple homogenous enzyme assay for carbonic anhydrase inside an agarose gel such that the enzyme in the gel would cause a color change throughout the gel. Thereafter, beads carrying combinatorial compounds via a photolinker were placed inside the gel and the compounds were partially released by UV-light. Compounds that inhibited the enzyme were observed as local zones of inhibition having less color change.

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Yet another example is described by Salmon. *et al.*, *Molecular Diversity* 2, 57-63 (1996). In this example, combinatorial libraries were screened for compounds that had cytotoxic effects on cancer cells growing in agar.

30 Another high throughput screening method is described in Beutel *et al.*, U.S. Patent 5,976,813. In this method, test samples are placed in a porous matrix. One or more

assay components are then placed within, on top of, or at the bottom of a matrix such as a gel, a plastic sheet, a filter, or other form of easily manipulated solid support. When samples are introduced to the porous matrix they diffuse sufficiently slowly, such that the assays can be performed without the test samples running together..

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Binding Assays

For binding assays, the test compound is preferably a small molecule that binds to and occupies, for example, the active site of the elongase HSEL01-like protein polypeptide, such that normal biological activity is prevented. Examples of such small molecules include, but are not limited to, small peptides or peptide-like molecules.

In binding assays, either the test compound or the elongase HSEL01-like protein polypeptide can comprise a detectable label, such as a fluorescent, radioisotopic, chemiluminescent, or enzymatic label, such as horseradish peroxidase, alkaline phosphatase, or luciferase. Detection of a test compound that is bound to the elongase HSEL01-like protein polypeptide can then be accomplished, for example, by direct counting of radioemission, by scintillation counting, or by determining conversion of an appropriate substrate to a detectable product.

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Alternatively, binding of a test compound to an elongase HSEL01-like protein polypeptide can be determined without labeling either of the interactants. For example, a microphysiometer can be used to detect binding of a test compound with an elongase HSEL01-like protein polypeptide. A microphysiometer (*e.g.*, Cytosensor™) is an analytical instrument that measures the rate at which a cell acidifies its environment using a light-addressable potentiometric sensor (LAPS). Changes in this acidification rate can be used as an indicator of the interaction between a test compound and an elongase HSEL01-like protein polypeptide (McConnell *et al.*, *Science* 257,

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and an elongase HSEL01-like protein polypeptide (McConnell *et al.*, *Science* 257, 1906-1912, 1992).

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Determining the ability of a test compound to bind to an elongase HSELO1-like protein polypeptide also can be accomplished using a technology such as real-time Bimolecular Interaction Analysis (BIA) (Sjolander & Urbanitzky, *Anal. Chem.* 63, 2338-2345, 1991, and Szabo *et al.*, *Curr. Opin. Struct. Biol.* 5, 699-705, 1995). BIA is a technology for studying biospecific interactions in real time, without labeling any of the interactants (e.g., BIACore™). Changes in the optical phenomenon surface plasmon resonance (SPR) can be used as an indication of real-time reactions between biological molecules.

encoding the protein that interacts with the elongase HSELO1-like protein polypeptide.

In yet another aspect of the invention, an elongases HSELO1-like protein polypeptide can be used as a "bait protein" in a two-hybrid assay or three-hybrid assay (see, e.g., U.S. Patent 5,283,317; Zervos *et al.*, *Cell* 72, 223-232, 1993; Madura *et al.*, *J. Biol. Chem.* 268, 12046-12054, 1993; Bartel *et al.*, *BioTechniques* 14, 920-924, 1993; Iwabuchi *et al.*, *Oncogene* 8, 1693-1696, 1993; and Brent W094/10300), to identify other proteins which bind to or interact with the elongase HSELO1-like protein polypeptide and modulate its activity.

The two-hybrid system is based on the modular nature of most transcription factors, which consist of separable DNA-binding and activation domains. Briefly, the assay utilizes two different DNA constructs. For example, in one construct, polynucleotide encoding an elongase HSELO1-like protein polypeptide can be fused to a polynucleotide encoding the DNA binding domain of a known transcription factor (e.g., GAL4). In the other construct a DNA sequence that encodes an unidentified protein ("prey" or "sample") can be fused to a polynucleotide that codes for the activation domain of the known transcription factor. If the "bait" and the "prey" proteins are able to interact *in vivo* to form an protein-dependent complex, the DNA-binding and activation domains of the transcription factor are brought into close proximity. This proximity allows transcription of a reporter gene (e.g., LacZ), which is operably linked to a transcriptional regulatory site responsive to the transcription factor. Expression of the reporter gene can be detected, and cell colonies containing the functional transcription factor can be isolated and used to obtain the DNA sequence

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encoding the protein that interacts with the elongase HSELO1-like protein polypeptide.

It may be desirable to immobilize either the elongase HSELO1-like protein polypeptide (or polynucleotide) or the test compound to facilitate separation of bound from unbound forms of one or both of the interactants, as well as to accommodate automation of the assay. Thus, either the elongase HSELO1-like protein polypeptide (or polynucleotide) or the test compound can be bound to a solid support. Suitable solid supports include, but are not limited to, glass or plastic slides, tissue culture plates, microtiter wells, tubes, silicon chips, or particles such as beads (including, but not limited to, latex, polystyrene, or glass beads). Any method known in the art can be used to attach the enzyme polypeptide (or polynucleotide) or test compound to a solid support, including use of covalent and non-covalent linkages, passive absorption, or pairs of binding moieties attached respectively to the polypeptide (or polynucleotide) or test compound and the solid support. Test compounds are preferably bound to the solid support in an array, so that the location of individual test compounds can be tracked. Binding of a test compound to an elongase HSELO1-like protein polypeptide (or polynucleotide) can be accomplished in any vessel suitable for containing the reactants. Examples of such vessels include microtiter plates, test tubes, and microcentrifuge tubes.

In one embodiment, the elongase HSELO1-like protein polypeptide is a fusion protein comprising a domain that allows the elongase HSELO1-like protein polypeptide to be bound to a solid support. For example, glutathione-S-transferase fusion proteins can be adsorbed onto glutathione sepharose beads (Sigma Chemical, St. Louis, Mo.) or glutathione derivatized microtiter plates, which are then combined with the test compound or the test compound and the non-adsorbed elongase HSELO1-like protein polypeptide; the mixture is then incubated under conditions conducive to complex formation (e.g., at physiological conditions for salt and pH).

Following incubation, the beads or microtiter plate wells are washed to remove any unbound components. Binding of the interactants can be determined either directly

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or indirectly, as described above. Alternatively, the complexes can be dissociated from the solid support before binding is determined.

Other techniques for immobilizing proteins or polynucleotides on a solid support also can be used in the screening assays of the invention. For example, either an elongase HSEL01-like protein polypeptide (or polynucleotide) or a test compound can be immobilized utilizing conjugation of biotin and streptavidin. Biotinylated elongase HSEL01-like protein polypeptides (or polynucleotides) or test compounds can be prepared from biotin-NHS(N-hydroxysuccinimide) using techniques well known in the art (e.g., biotinylation kit, Pierce Chemicals, Rockford, IL) and immobilized in the wells of streptavidin-coated 96 well plates (Pierce Chemical). Alternatively, antibodies which specifically bind to an elongase HSEL01-like protein polypeptide, polynucleotide, or a test compound, but which do not interfere with a desired binding site, such as the active site of the elongase HSEL01-like protein polypeptide, can be derivatized to the wells of the plate. Unbound target or protein can be trapped in the wells by antibody conjugation.

Methods for detecting such complexes, in addition to those described above for the GST-immobilized complexes, include immunodetection of complexes using antibodies which specifically bind to the elongase HSEL01-like protein polypeptide or test compound, enzyme-linked assays which rely on detecting an activity of the elongase HSEL01-like protein polypeptide, and SDS gel electrophoresis under non-reducing conditions.

Screening for test compounds which bind to an elongase HSEL01-like protein polypeptide or polynucleotide also can be carried out in an intact cell. Any cell which comprises an elongase HSEL01-like protein polypeptide or polynucleotide can be used in a cell-based assay system. An elongase HSEL01-like protein polynucleotide can be naturally occurring in the cell, or can be introduced using techniques such as those described above. Binding of the test compound to an elongase

HSEL01-like protein polypeptide or polynucleotide is determined as described above.

Enzyme Assays

5 Test compounds can be tested for the ability to increase or decrease the enzyme activity of a human elongase HSEL01-like protein polypeptide. Enzyme activity can be measured, for example, as described in Ghioni *et al.*, *Biochim Biophys Acta* 1999 Feb 25;1437(2):170-81; Tocher, *Lipids* 1993 Apr;28(4):267-72; or Chang *et al.*, J 10 Nutr 1992 Nov;122(11):2074-80.

15 Enzyme assays can be carried out after contacting either a purified elongase HSEL01-like protein polypeptide, a cell membrane preparation, or an intact cell with a test compound. A test compound that decreases an elongase activity of an elongase HSEL01-like protein polypeptide by at least about 10, preferably about 50, more preferably about 75, 90, or 100% is identified as a potential therapeutic agent for decreasing elongase HSEL01-like protein activity. A test compound which increases an elongase activity of a human elongase HSEL01-like protein polypeptide by at least about 10, preferably about 50, more preferably about 75, 90, or 100% is identified as a potential therapeutic agent for increasing human elongase HSEL01-like protein activity.

20 Gene Expression

25 In another embodiment, test compounds that increase or decrease elongase HSEL01-like protein gene expression are identified. An elongase HSEL01-like protein polynucleotide is contacted with a test compound, and the expression of an RNA or polypeptide product of the elongase HSEL01-like protein polynucleotide is determined. The level of expression of appropriate mRNA or polypeptide in the presence of the test compound is compared to the level of expression of mRNA or polypeptide in the absence of the test compound. The test compound can then be identified as a

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modulator of expression based on this comparison. For example, when expression of mRNA or polypeptide is greater in the presence of the test compound than in its absence, the test compound is identified as a stimulator or enhancer of the mRNA or polypeptide expression. Alternatively, when expression of the mRNA or polypeptide is less in the presence of the test compound than in its absence, the test compound is identified as an inhibitor of the mRNA or polypeptide expression.

The level of elongase HSEL01-like protein mRNA or polypeptide expression in the cells can be determined by methods well known in the art for detecting mRNA or polypeptide. Either qualitative or quantitative methods can be used. The presence of polypeptide products of an elongase HSEL01-like protein polynucleotide can be determined, for example, using a variety of techniques known in the art, including immunochemical methods such as radioimmunoassay, Western blotting, and immunohistochemistry. Alternatively, polypeptide synthesis can be determined *in vivo*, in a cell culture, or in an *in vitro* translation system by detecting incorporation of labeled amino acids into an elongase HSEL01-like protein polypeptide.

Such screening can be carried out either in a cell-free assay system or in an intact cell. Any cell that expresses an elongase HSEL01-like protein polynucleotide can be used in a cell-based assay system. The elongase HSEL01-like protein polynucleotide can be naturally occurring in the cell or can be introduced using techniques such as those described above. Either a primary culture or an established cell line, such as CHO or human embryonic kidney 293 cells, can be used.

25 Pharmaceutical Compositions
The invention also provides pharmaceutical compositions that can be administered to a patient to achieve a therapeutic effect. Pharmaceutical compositions of the invention can comprise, for example, an elongase HSEL01-like protein polypeptide, elongase HSEL01-like protein polynucleotide, ribozymes or antisense oligonucleotides, antibodies which specifically bind to an elongase HSEL01-like protein poly-

peptide, or mimetics, activators, or inhibitors of an elongase HSEL01-like protein polypeptide activity. The compositions can be administered alone or in combination with at least one other agent, such as stabilizing compound, which can be administered in any sterile, biocompatible pharmaceutical carrier, including, but not limited to, saline, buffered saline, dextrose, and water. The compositions can be administered to a patient alone, or in combination with other agents, drugs or hormones.

In addition to the active ingredients, these pharmaceutical compositions can contain suitable pharmaceutically acceptable carriers comprising excipients and auxiliaries that facilitate processing of the active compounds into preparations which can be used pharmaceutically. Pharmaceutical compositions of the invention can be administered by any number of routes including, but not limited to, oral, intravenous, intramuscular, intra-arterial, intramedullary, intrathecal, intraventricular, transdermal, subcutaneous, intraperitoneal, intranasal, parenteral, topical, sublingual, or rectal means. Pharmaceutical compositions for oral administration can be formulated using pharmaceutically acceptable carriers well known in the art in dosages suitable for oral administration. Such carriers enable the pharmaceutical compositions to be formulated as tablets, pills, dragees, capsules, liquids, gels, syrups, slurries, suspensions, and the like, for ingestion by the patient.

10 **15** Pharmaceutical preparations for oral use can be obtained through combination of active compounds with solid excipient, optionally grinding a resulting mixture, and processing the mixture of granules, after adding suitable auxiliaries, if desired, to obtain tablets or dragee cores. Suitable excipients are carbohydrate or protein fillers, such as sugars, including lactose, sucrose, mannitol, or sorbitol; starch from corn, wheat, rice, potato, or other plants; cellulose, such as methyl cellulose, hydroxypropylmethyl-cellulose, or sodium carboxymethyl/cellulose; gums including arabic and tragacanth; and proteins such as gelatin and collagen. If desired, disintegrating or solubilizing agents can be added, such as the cross-linked polyvinyl pyrrolidone, agar, alginic acid, or a salt thereof, such as sodium alginate.

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5 Dragee cores can be used in conjunction with suitable coatings, such as concentrated sugar solutions, which also can contain gum arabic, talc, polyvinylpyrrolidone, carbopol gel, polyethylene glycol, and/or titanium dioxide, lacquer solutions, and suitable organic solvents or solvent mixtures. Dye stuffs or pigments can be added to the tablets or dragee coatings for product identification or to characterize the quantity of active compound, i.e., dosage.

10 Pharmaceutical preparations that can be used orally include push-fit capsules made of gelatin, as well as soft, sealed capsules made of gelatin and a coating, such as glycerol or sorbitol. Push-fit capsules can contain active ingredients mixed with a filler or binders, such as lactose or starches, lubricants, such as talc or magnesium stearate, and, optionally, stabilizers. In soft capsules, the active compounds can be dissolved or suspended in suitable liquids, such as fatty oils, liquid, or liquid polyethylene glycol with or without stabilizers.

15 Pharmaceutical formulations suitable for parenteral administration can be formulated in aqueous solutions, preferably in physiologically compatible buffers such as Hanks' solution, Ringer's solution, or physiologically buffered saline. Aqueous injection suspensions can contain substances that increase the viscosity of the suspension, such as sodium carboxymethyl cellulose, sorbitol, or dextran. Additionally, suspensions of the active compounds can be prepared as appropriate oily injection suspensions. Suitable lipophilic solvents or vehicles include fatty oils such as sesame oil, or synthetic fatty acid esters, such as ethyl oleate or triglycerides, or liposomes. Non-lipid polycationic amino polymers also can be used for delivery. Optionally, the suspension also can contain suitable stabilizers or agents that increase the solubility of the compounds to allow for the preparation of highly concentrated solutions. For topical or nasal administration, penetrants appropriate to the particular barrier to be permeated are used in the formulation. Such penetrants are generally known in the art.

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20 The pharmaceutical compositions of the present invention can be manufactured in a manner that is known in the art, e.g., by means of conventional mixing, dissolving, granulating, dragee-making, levigating, emulsifying, encapsulating, entrapping, or lyophilizing processes. The pharmaceutical composition can be provided as a salt and can be formed with many acids, including but not limited to, hydrochloric, sulfuric, acetic, lactic, tartaric, malic, succinic, etc. Salts tend to be more soluble in aqueous or other protonic solvents than are the corresponding free base forms. In other cases, the preferred preparation can be a lyophilized powder which can contain any or all of the following: 1-50 mM histidine, 0.1%-2% sucrose, and 2-7% mannitol, at a pH range of 4.5 to 5.5, that is combined with buffer prior to use.

25 Further details on techniques for formulation and administration can be found in the latest edition of REMMINGTON'S PHARMACEUTICAL SCIENCES (Maack Publishing Co., Easton, Pa.). After pharmaceutical compositions have been prepared, they can be placed in an appropriate container and labeled for treatment of an indicated condition. Such labeling would include amount, frequency, and method of administration.

30 Therapeutic Indications and Methods

35 Diabetes

40 Diabetes mellitus is a common metabolic disorder characterized by an abnormal elevation in blood glucose, alterations in lipids and abnormalities (complications) in the cardiovascular system, eye, kidney and nervous system. Diabetes is divided into two separate diseases: type 1 diabetes (juvenile onset), which results from a loss of cells which make and secrete insulin, and type 2 diabetes (adult onset), which is caused by a defect in insulin secretion and a defect in insulin action.

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Type I diabetes is initiated by an autoimmune reaction that attacks the insulin secreting cells (beta cells) in the pancreatic islets. Agents that prevent this reaction from occurring or that stop the reaction before destruction of the beta cells has been accomplished are potential therapies for this disease. Other agents that induce beta cell proliferation and regeneration also are potential therapies.

Type II diabetes is the most common of the two diabetic conditions (6% of the population). The defect in insulin secretion is an important cause of the diabetic condition and results from an inability of the beta cell to properly detect and respond to rises in blood glucose levels with insulin release. Therapies that increase the response by the beta cell to glucose would offer an important new treatment for this disease.

The defect in insulin action in Type II diabetic subjects is another target for therapeutic intervention. Agents that increase the activity of the insulin receptor in muscle, liver, and fat will cause a decrease in blood glucose and a normalization of plasma lipids. The receptor activity can be increased by agents that directly stimulate the receptor or that increase the intracellular signals from the receptor. Other therapies can directly activate the cellular end process, i.e. glucose transport or various enzyme systems, to generate an insulin-like effect and therefore a produce beneficial outcome. Because overweight subjects have a greater susceptibility to Type II diabetes, any agent that reduces body weight is a possible therapy.

Both Type I and Type diabetes can be treated with agents that mimic insulin action or that treat diabetic complications by reducing blood glucose levels. Likewise, agents that reduces new blood vessel growth can be used to treat the eye complications that develop in both diseases.

Cancer

Cancer is a disease fundamentally caused by oncogenic cellular transformation. There are several hallmarks of transformed cells that distinguish them from their normal counterparts and underlie the pathophysiology of cancer. These include uncontrolled cellular proliferation, unresponsiveness to normal death-inducing signals (immortalization), increased cellular motility and invasiveness, increased ability to recruit blood supply through induction of new blood vessel formation (angiogenesis), genetic instability, and dysregulated gene expression. Various combinations of these aberrant physiologies, along with the acquisition of drug-resistance frequently lead to an intractable disease state in which organ failure and patient death ultimately ensue.

Most standard cancer therapies target cellular proliferation and rely on the differential proliferative capacities between transformed and normal cells for their efficacy. This approach is hindered by the facts that several important normal cell types are also highly proliferative and that cancer cells frequently become resistant to these agents. Thus, the therapeutic indices for traditional anti-cancer therapies rarely exceed 2.0.

The advent of genomics-driven molecular target identification has opened up the possibility of identifying new cancer-specific targets for therapeutic intervention that will provide safer, more effective treatments for cancer patients. Thus, newly discovered tumor-associated genes and their products can be tested for their role(s) in disease and used as tools to discover and develop innovative therapies. Genes playing important roles in any of the physiological processes outlined above can be characterized as cancer targets.

Genes or gene fragments identified through genomics can readily be expressed in one or more heterologous expression systems to produce functional recombinant proteins. These proteins are characterized *in vitro* for their biochemical properties and then used as tools in high-throughput molecular screening programs to identify chemical

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modulators of their biochemical activities. Agonists and/or antagonists of target protein activity can be identified in this manner and subsequently tested in cellular and *in vivo* disease models for anti-cancer activity. Optimization of lead compounds with iterative testing in biological models and detailed pharmacokinetic and toxicological analyses form the basis for drug development and subsequent testing in humans.

CNS disorders

10 CNS disorders which may be treated include brain injuries, cerebrovascular diseases and their consequences, Parkinson's disease, corticobasal degeneration, motor neuron disease, dementia, including ALS, multiple sclerosis, traumatic brain injury, stroke, post-stroke, post-traumatic brain injury, and small-vessel cerebrovascular disease. Dementias, such as Alzheimer's disease, vascular dementia, dementia with Lewy bodies, frontotemporal dementia and Parkinsonism linked to chromosome 17, frontotemporal dementias, including Pick's disease, progressive nuclear palsy, corticobasal degeneration, Huntington's disease, thalamic degeneration, Creutzfeld-Jakob dementia, HIV dementia, schizophrenia with dementia, and Korsakoff's psychosis also can be treated. Similarly, it may be possible to treat cognitive-related disorders, such as mild cognitive impairment, age-associated memory impairment, age-related cognitive decline, vascular cognitive impairment, attention deficit disorders, attention deficit hyperactivity disorders, and memory disturbances in children with learning disabilities, by regulating the activity of human elongase HSELO1-like protein.

25 Pain that is associated with CNS disorders also can be treated by regulating the activity of human elongase HSELO1-like protein. Pain which can be treated includes that associated with central nervous system disorders, such as multiple sclerosis, spinal cord injury, sciatica, failed back surgery syndrome, traumatic brain injury, epilepsy, Parkinson's disease, post-stroke, and vascular lesions in the brain and spinal cord (e.g., infarct, hemorrhage, vascular malformation). Non-central neuro-

pathic pain includes that associated with post mastectomy pain, reflex sympathetic dystrophy (RSD), trigeminal neuralgioradiculargia, post-surgical pain, HTV/AIDS related pain, cancer pain, metabolic neuropathies (e.g., diabetic neuropathy, vasculitic neuropathy secondary to connective tissue disease), paraneoplastic polyneuropathy associated, for example, with carcinoma of lung, or leukemia, or lymphoma, or carcinoma of prostate, colon or stomach, trigeminal neuralgia, cranial neuralgias, and post-herpetic neuralgia. Pain associated with cancer and cancer treatment also can be treated, as can headache pain (for example, migraine with aura, migraine without aura, and other migraine disorders), episodic and chronic tension-type headache, tension-type like headache, cluster headache, and chronic paroxysmal hemicrania.

This invention further pertains to the use of novel agents identified by the screening assays described above. Accordingly, it is within the scope of this invention to use a test compound identified as described herein in an appropriate animal model. For example, an agent identified as described herein (e.g., a modulating agent, an antisense nucleic acid molecule, a specific antibody, ribozyme, or an elongase HSELO1-like protein polypeptide binding molecule) can be used in an animal model to determine the efficacy, toxicity, or side effects of treatment with such an agent. Alternatively, an agent identified as described herein can be used in an animal model to determine the mechanism of action of such an agent. Furthermore, this invention pertains to uses of novel agents identified by the above-described screening assays for treatments as described herein.

A reagent which affects elongase HSELO1-like protein activity can be administered to a human cell, either *in vitro* or *in vivo*, to reduce elongase HSELO1-like protein activity. The reagent preferably binds to an expression product of a human elongase HSELO1-like protein gene. If the expression product is a protein, the reagent is preferably an antibody. For treatment of human cells *ex vivo*, an antibody can be added to a preparation of stem cells that have been removed from the body. The cells can then be replaced in the same or another human body, with or without clonal propagation, as is known in the art.

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In one embodiment, the reagent is delivered using a liposome. Preferably, the liposome is stable in the animal into which it has been administered for at least about 30 minutes, more preferably for at least about 1 hour, and even more preferably for at least about 24 hours. A liposome comprises a lipid composition that is capable of targeting a reagent, particularly a polynucleotide, to a particular site in an animal, such as a human. Preferably, the lipid composition of the liposome is capable of targeting to a specific organ of an animal, such as the lung, liver, spleen, heart, brain, lymph nodes, and skin.

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A liposome useful in the present invention comprises a lipid composition that is capable of fusing with the plasma membrane of the targeted cell to deliver its contents to the cell. Preferably, the transfection efficiency of a liposome is about 0.5 μg of DNA per 16 nmole of liposome delivered to about 10^6 cells, more preferably about 1.0 μg of DNA per 16 nmole of liposome delivered to about 10^6 cells, and even more preferably about 2.0 μg of DNA per 16 nmol of liposome delivered to about 10^6 cells. Preferably, a liposome is between about 100 and 500 nm, more preferably between about 150 and 450 nm, and even more preferably between about 200 and 400 nm in diameter.

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Suitable liposomes for use in the present invention include those liposomes standardly used in, for example, gene delivery methods known to those of skill in the art. More preferred liposomes include liposomes having a polycationic lipid composition and/or liposomes having a cholesterol backbone conjugated to polyethylene glycol. Optionally, a liposome comprises a compound capable of targeting the liposome to a particular cell type, such as a cell-specific ligand exposed on the outer surface of the liposome.

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Complexing a liposome with a reagent such as an antisense oligonucleotide or ribozyme can be achieved using methods that are standard in the art (see, for example, U.S. Patent 5,705,151). Preferably, from about 0.1 μg to about 10 μg of

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polynucleotide is combined with about 8 nmol of liposomes, more preferably from about 0.5 μg to about 5 μg of polynucleotides are combined with about 8 nmol liposomes, and even more preferably about 1.0 μg of polynucleotides is combined with about 8 nmol liposomes.

5 In another embodiment, antibodies can be delivered to specific tissues *in vivo* using receptor-mediated targeted delivery. Receptor-mediated DNA delivery techniques are taught in, for example, Findeis *et al.*, *Trends in Biotechnol.* 11, 202-05 (1993); Chiou *et al.*, *GENE THERAPEUTICS: METHODS AND APPLICATIONS OF DIRECT GENE TRANSFER* (J.A. Wolff, ed.) (1994); Wu & Wu, *J. Biol. Chem.* 263, 621-24 (1988); Wu *et al.*, *J. Biol. Chem.* 269, 542-46 (1994); Zenke *et al.*, *Proc. Natl. Acad. Sci. U.S.A.* 87, 3655-59 (1990); Wu *et al.*, *J. Biol. Chem.* 266, 338-42 (1991).

Determination of a Therapeutically Effective Dose

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The determination of a therapeutically effective dose is well within the capability of those skilled in the art. A therapeutically effective dose refers to that amount of active ingredient which increases or decreases elongase HSEL01-like protein activity relative to the elongase HSEL01-like protein activity which occurs in the absence of the therapeutically effective dose.

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For any compound, the therapeutically effective dose can be estimated initially either in cell culture assays or in animal models, usually mice, rabbits, dogs, or pigs. The animal model also can be used to determine the appropriate concentration range and route of administration. Such information can then be used to determine useful doses and routes for administration in humans.

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Therapeutic efficacy and toxicity, e.g., ED₅₀ (the dose therapeutically effective in 50% of the population) and LD₅₀ (the dose lethal to 50% of the population), can be determined by standard pharmaceutical procedures in cell cultures or experimental

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animals. The dose ratio of toxic to therapeutic effects is the therapeutic index, and it can be expressed as the ratio, LD₅₀/ED₅₀.

Pharmaceutical compositions that exhibit large therapeutic indices are preferred. The data obtained from cell culture assays and animal studies is used in formulating a range of dosage for human use. The dosage contained in such compositions is preferably within a range of circulating concentrations that include the ED₅₀ with little or no toxicity. The dosage varies within this range depending upon the dosage form employed, sensitivity of the patient, and the route of administration.

The exact dosage will be determined by the practitioner, in light of factors related to the subject that requires treatment. Dosage and administration are adjusted to provide sufficient levels of the active ingredient or to maintain the desired effect. Factors that can be taken into account include the severity of the disease state, general health of the subject, age, weight, and gender of the subject, diet, time and frequency of administration, drug combination(s), reaction sensitivities, and tolerance/response to therapy. Long-acting pharmaceutical compositions can be administered every 3 to 4 days, every week, or once every two weeks depending on the half-life and clearance rate of the particular formulation.

Normal dosage amounts can vary from 0.1 to 100,000 micrograms, up to a total dose of about 1 g, depending upon the route of administration. Guidance as to particular dosages and methods of delivery is provided in the literature and generally available to practitioners in the art. Those skilled in the art will employ different formulations for nucleotides than for proteins or their inhibitors. Similarly, delivery of polynucleotides or polypeptides will be specific to particular cells, conditions, locations, etc.

If the reagent is a single-chain antibody, polynucleotides encoding the antibody can be constructed and introduced into a cell either *ex vivo* or *in vivo* using well-established techniques including, but not limited to, transfecting-polycation-mediated

DNA transfer, transfection with naked or encapsulated nucleic acids, liposome-mediated cellular fusion, intracellular transportation of DNA-coated latex beads, protoplast fusion, viral infection, electroporation, "gene gun," and DEAE- or calcium phosphate-mediated transfection.

Effective *in vivo* dosages of an antibody are in the range of about 5 µg to about 50 µg/kg, about 50 µg to about 5 mg/kg, about 100 µg to about 500 µg/kg of patient body weight, and about 200 to about 250 µg/kg of patient body weight. For administration of polynucleotides encoding single-chain antibodies, effective *in vivo*

dosages are in the range of about 100 ng to about 200 ng, 500 ng to about 50 mg, about 1 µg to about 2 mg, about 5 µg to about 500 µg, and about 20 µg to about 100 µg of DNA.

If the expression product is mRNA, the reagent is preferably an antisense oligonucleotide or a ribozyme. Polynucleotides that express antisense oligonucleotides or ribozymes can be introduced into cells by a variety of methods, as described above.

Preferably, a reagent reduces expression of an elongase HSEL01-like protein gene or the activity of an elongase HSEL01-like protein polypeptide by at least about 10, preferably about 50, more preferably about 75, 90, or 100% relative to the absence of the reagent. The effectiveness of the mechanism chosen to decrease the level of expression of an elongase HSEL01-like protein gene or the activity of an elongase HSEL01-like protein polypeptide can be assessed using methods well known in the art, such as hybridization of nucleotide probes to elongase HSEL01-like protein-specific mRNA, quantitative RT-PCR, immunologic detection of an elongase HSEL01-like protein polypeptide, or measurement of elongase HSEL01-like protein activity.

In any of the embodiments described above, any of the pharmaceutical compositions of the invention can be administered in combination with other appropriate therapeutic agents. Selection of the appropriate agents for use in combination therapy can

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DNA transfer, transfection with naked or encapsulated nucleic acids, liposome-mediated cellular fusion, intracellular transportation of DNA-coated latex beads, protoplast fusion, viral infection, electroporation, "gene gun," and DEAE- or calcium phosphate-mediated transfection.

Effective *in vivo* dosages of an antibody are in the range of about 5 µg to about 50 µg/kg, about 50 µg to about 5 mg/kg, about 100 µg to about 500 µg/kg of patient body weight, and about 200 to about 250 µg/kg of patient body weight. For administration of polynucleotides encoding single-chain antibodies, effective *in vivo*

dosages are in the range of about 100 ng to about 200 ng, 500 ng to about 50 mg, about 1 µg to about 2 mg, about 5 µg to about 500 µg, and about 20 µg to about 100 µg of DNA.

If the expression product is mRNA, the reagent is preferably an antisense oligonucleotide or a ribozyme. Polynucleotides that express antisense oligonucleotides or ribozymes can be introduced into cells by a variety of methods, as described above.

Preferably, a reagent reduces expression of an elongase HSEL01-like protein gene or the activity of an elongase HSEL01-like protein polypeptide by at least about 10, preferably about 50, more preferably about 75, 90, or 100% relative to the absence of the reagent. The effectiveness of the mechanism chosen to decrease the level of expression of an elongase HSEL01-like protein gene or the activity of an elongase HSEL01-like protein polypeptide can be assessed using methods well known in the art, such as hybridization of nucleotide probes to elongase HSEL01-like protein-specific mRNA, quantitative RT-PCR, immunologic detection of an elongase HSEL01-like protein polypeptide, or measurement of elongase HSEL01-like protein activity.

In any of the embodiments described above, any of the pharmaceutical compositions of the invention can be administered in combination with other appropriate therapeutic agents. Selection of the appropriate agents for use in combination therapy can

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be made by one of ordinary skill in the art, according to conventional pharmaceutical principles. The combination of therapeutic agents can act synergistically to effect the treatment or prevention of the various disorders described above. Using this approach, one may be able to achieve therapeutic efficacy with lower dosages of each agent, thus reducing the potential for adverse side effects.

Any of the therapeutic methods described above can be applied to any subject in need of such therapy, including, for example, mammals such as dogs, cats, cows, horses, rabbits, monkeys, and most preferably, humans.

Diagnostic Methods

Human elongase HSEL01-like protein also can be used in diagnostic assays for detecting diseases and abnormalities or susceptibility to diseases and abnormalities related to the presence of mutations in the nucleic acid sequences that encode the enzyme. For example, differences can be determined between the cDNA or genomic sequence encoding elongase HSEL01-like protein in individuals afflicted with a disease and in normal individuals. If a mutation is observed in some or all of the afflicted individuals but not in normal individuals, then the mutation is likely to be the causative agent of the disease.

Sequence differences between a reference gene and a gene having mutations can be revealed by the direct DNA sequencing method. In addition, cloned DNA segments can be employed as probes to detect specific DNA segments. The sensitivity of this method is greatly enhanced when combined with PCR. For example, a sequencing primer can be used with a double-stranded PCR product or a single-stranded template molecule generated by a modified PCR. The sequence determination is performed by conventional procedures using radiolabeled nucleotides or by automatic sequencing procedures using fluorescent tags.

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Genetic testing based on DNA sequence differences can be carried out by detection of alteration in electrophoretic mobility of DNA fragments in gels with or without denaturing agents. Small sequence deletions and insertions can be visualized, for example, by high resolution gel electrophoresis. DNA fragments of different sequences can be distinguished on denaturing formamide gradient gels in which the mobilities of different DNA fragments are retarded in the gel at different positions according to their specific melting or partial melting temperatures (see, e.g., Myers *et al.*, *Science* 230, 1242, 1985). Sequence changes at specific locations can also be revealed by nuclease protection assays, such as RNase and S 1 protection or the chemical cleavage method (e.g., Cotton *et al.*, *Proc. Natl. Acad. Sci. USA* 85, 4397-4401, 1985). Thus, the detection of a specific DNA sequence can be performed by methods such as hybridization, RNase protection, chemical cleavage, direct DNA sequencing or the use of restriction enzymes and Southern blotting of genomic DNA. In addition to direct methods such as gel-electrophoresis and DNA sequencing, mutations can also be detected by *in situ* analysis.

Altered levels of elongase HSEL01-like protein also can be detected in various tissues. Assays used to detect levels of the receptor polypeptides in a body sample, such as blood or a tissue biopsy, derived from a host are well known to those of skill in the art and include radioimmunoassays, competitive binding assays, Western blot analysis, and ELISA assays.

All patents and patent applications cited in this disclosure are expressly incorporated herein by reference. The above disclosure generally describes the present invention.

A more complete understanding can be obtained by reference to the following specific examples, which are provided for purposes of illustration only and are not intended to limit the scope of the invention.

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EXAMPLE 1***Detection of elongase HSELO1-like protein activity***

5 The polynucleotide of SEQ ID NO: 1 is inserted into the expression vector pCEV4 and the expression vector pCEV4-elongase HSELO1-like protein polypeptide obtained is transfected into human embryonic kidney 293 cells. From these cells extracts are obtained and elongase activity is measured in a volume of 200 µl containing 50 mM Tris (pH 7.5), 1 mM MgCl₂, 150µM Triton X-100, 1 mM NADPH, 1 mM NADH, 10 mM betamercaptoethanol, 40 µM acyl-CoA acceptor (either palmitoyl-CoA, stearoyl-CoA, or eicosanoyl-CoA), and 60 µM [2-14] malonyl-CoA (0.05 µCi/ml) at 37°C. The reaction is initiated by the addition of 0.3 to 1.0 mg of the cell extract. Protein concentrations are determined using the Bio-rad protein assay reagent (Bio-Laboratories, Hercules, Calif.). At various times, the reaction is terminated by adding 200µl of 5 M KOH-10% methanol MeOH and heating at 80°C for 1 h. Following addition of 200 µl of 10 N H₂SO₄, fatty acids are recovered by two 1.5-ml extractions into hexane. The extracted fatty acids are resolved by silica gel TLC using hexane-diethyl ether-acetic (30:70:1) as the developing solvent. The radiolabeled fatty acids are detected and quantified using a PhosphorImager SI (Molecular Dynamics, Inc., Sunnyvale, Calif.). It is shown that the polypeptide of SEQ ID NO: 2 has a elongase HSELO1-like protein activity.

EXAMPLE 2***Expression of recombinant human elongase HSELO1-like protein***

25 The *Pichia pastoris* expression vector pPICZB (Invitrogen, San Diego, CA) is used to produce large quantities of recombinant human elongase HSELO1-like protein polypeptides in yeast. The elongase HSELO1-like protein-encoding DNA sequence is derived from SEQ ID NO: 1. Before insertion into vector pPICZB, the DNA sequence is modified by well known methods in such a way that it contains at its

5'-end an initiation codon and at its 3'-end an enterokinase cleavage site, a His6 reporter tag and a termination codon. Moreover, at both termini recognition sequences for restriction endonucleases are added and after digestion of the multiple cloning site of pPICZB with the corresponding restriction enzymes the modified DNA sequence is ligated into pPICZB. This expression vector is designed for inducible expression in *Pichia pastoris*, driven by a yeast promoter. The resulting pPICZB/md-His6 vector is used to transform the yeast.

The yeast is cultivated under usual conditions in 5 liter shake flasks and the recombinantly produced protein isolated from the culture by affinity chromatography (Ni-NTA-Resin) in the presence of 8 M urea. The bound polypeptide is eluted with buffer, pH 3.5, and neutralized. Separation of the polypeptide from the His6 reporter tag is accomplished by site-specific proteolysis using enterokinase (Invitrogen, San Diego, CA) according to manufacturer's instructions. Purified human elongase HSELO1-like protein polypeptide is obtained.

EXAMPLE 3

Identification of test compounds that bind to elongase HSELO1-like protein polypeptides

10 Purified elongase HSELO1-like protein polypeptides comprising a glutathione-S-transferase protein and absorbed onto glutathione-derivatized wells of 96-well microtiter plates are contacted with test compounds from a small molecule library at pH 7.0 in a physiological buffer solution. Human elongase HSELO1-like protein polypeptides comprise the amino acid sequence shown in SEQ ID NO: 2. The test compounds comprise a fluorescent tag. The samples are incubated for 5 minutes to one hour. Control samples are incubated in the absence of a test compound.

15 The buffer solution containing the test compounds is washed from the wells.

20 Binding of a test compound to an elongase HSELO1-like protein polypeptide is

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detected by fluorescence measurements of the contents of the wells. A test compound that increases the fluorescence in a well by at least 15% relative to fluorescence of a well in which a test compound is not incubated is identified as a compound which binds to an elongase HSEL01-like protein polypeptide.

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EXAMPLE 4

Identification of a test compound which decreases elongase HSEL01-like protein gene expression

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A test compound is administered to a culture of human cells transfected with an elongase HSEL01-like protein expression construct and incubated at 37°C for 10 to 45 minutes. A culture of the same type of cells that have not been transfected is incubated for the same time without the test compound to provide a negative control.

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RNA is isolated from the two cultures as described in Chirgwin *et al.*, *Biochem. 18*, 5294-99, 1979). Northern blots are prepared using 20 to 30 µg total RNA and hybridized with a ³²P-labeled elongase HSEL01-like protein-specific probe at 65°C in Express-hyb (CLONTECH). The probe comprises at least 11 contiguous nucleotides selected from the complement of SEQ ID NO: 1. A test compound that decreases the elongase HSEL01-like protein-specific signal relative to the signal obtained in the absence of the test compound is identified as an inhibitor of elongase HSEL01-like protein gene expression.

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EXAMPLE 5

Identification of a test compound which decreases elongase HSEL01-like protein activity

A test compound is administered to a culture of human cells transfected with an elongase HSEL01-like protein expression construct and incubated at 37 °C for 10 to

45 minutes. A culture of the same type of cells that have not been transfected is incubated for the same time without the test compound to provide a negative control. elongase HSEL01-like protein activity is measured using a method of Ghioni *et al.*, *Biochim. Biophys. Acta 1999 Feb 25;1437(2):170-81; Tocher, Lipids 1993 Apr;28(4):267-72; or Chang *et al.*, J Nutr 1992 Nov;122(11):2074-80.*

EXAMPLE 4

A test compound which decreases the elongase HSEL01-like protein activity of the elongase HSEL01-like protein relative to the elongase HSEL01-like protein activity in the absence of the test compound is identified as an inhibitor of elongase HSEL01-like protein activity.

EXAMPLE 6

Tissue-specific expression of elongase HSEL01-like protein

The qualitative expression pattern of elongase HSEL01-like protein in various tissues is determined by Reverse Transcription-Polymerase Chain Reaction (RT-PCR).

To demonstrate that elongase HSEL01-like protein is involved in CNS disorders, the following tissues are screened: fetal and adult brain, muscle, heart, lung, kidney, liver, thymus, testis, colon, placenta, trachea, pancreas, kidney, gastric mucosa, colon, liver, cerebellum, skin, cortex (Alzheimer's and normal), hypothalamus, cortex, amygdala, cerebellum, hippocampus, choroid, plexus, thalamus, and spinal cord.

To demonstrate that elongase HSEL01-like protein is involved in cancer, expression is determined in the following tissues: adrenal gland, bone marrow, brain, cerebellum, colon, fetal brain, fetal liver, heart, kidney, liver, lung, mammary gland, pancreas, placenta, prostate, salivary gland, skeletal muscle, small intestine, spinal cord, spleen, stomach, testis, thymus, thyroid, trachea, uterus, and peripheral blood

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lymphocytes. Expression in the following cancer cell lines also is determined: DU-145 (prostate), NCI-H125 (lung), HT-29 (colon), COLO-205 (colon), A-549 (lung), NCI-H460 (lung), HT-116 (colon), DLD-1 (colon), MDA-MD-231 (breast), LS174T (colon), ZF-75 (breast), MDA-MN-435 (breast), HT-1080, MCF-7 (breast), and U87. Matched pairs of malignant and normal tissue from the same patient also are tested.

To demonstrate that elongase HSELO1-like protein is involved in the disease process of diabetes, the following whole body panel is screened to show predominant or relatively high expression: subcutaneous and mesenteric adipose tissue, adrenal gland, bone marrow, brain, colon, fetal brain, heart, hypothalamus, kidney, liver, lung, mammary gland, pancreas, placenta, prostate, salivary gland, skeletal muscle, small intestine, spleen, stomach, testis, thymus, thyroid, trachea, and uterus. Human islet cells and an islet cell library also are tested. As a final step, the expression of elongase HSELO1-like protein in cells derived from normal individuals with the expression of cells derived from diabetic individuals is compared.

Quantitative expression profiling. Quantitative expression profiling is performed by the form of quantitative PCR analysis called "kinetic analysis" firstly described in Higuchi *et al.*, *BioTechnology* 10, 413-17, 1992, and Higuchi *et al.*, *BioTechnology* 11, 1026-30, 1993. The principle is that at any given cycle within the exponential phase of PCR, the amount of product is proportional to the initial number of template copies.

If the amplification is performed in the presence of an internally quenched fluorescent oligonucleotide (TaqMan probe) complementary to the target sequence, the probe is cleaved by the 5'-3' endonuclease activity of Taq DNA polymerase and a fluorescent dye released in the medium (Holland *et al.*, *Proc. Natl. Acad. Sci. U.S.A.* 88, 7276-80, 1991). Because the fluorescence emission will increase in direct proportion to the amount of the specific amplified product, the exponential growth phase of PCR product can be detected and used to determine the initial template

concentration (Heid *et al.*, *Genome Res.* 6, 986-94, 1996, and Gibson *et al.*, *Genome Res.* 6, 995-1001, 1996).

The amplification of an endogenous control can be performed to standardize the amount of sample RNA added to a reaction. In this kind of experiment, the control of choice is the 18S ribosomal RNA. Because reporter dyes with differing emission spectra are available, the target and the endogenous control can be independently quantified in the same tube if probes labeled with different dyes are used.

All "real time PCR" measurements of fluorescence are made in the ABI Prism 7700. *RNA extraction and cDNA preparation.* Total RNA from the tissues listed above are used for expression quantification. RNAs labeled "from autopsy" were extracted from autopic tissues with the TRIzol reagent (Life Technologies, MD) according to the manufacturer's protocol.

Fifty µg of each RNA were treated with DNase I for 1 hour at 37°C in the following reaction mix: 0.2 U/µl RNase-free DNase I (Roche Diagnostics, Germany); 0.4 U/µl RNase inhibitor (PE Applied Biosystems, CA); 10 mM Tris-HCl pH 7.9; 10 mM MgCl₂; 50 mM NaCl; and 1 mM DTT.

After incubation, RNA is extracted once with 1 volume of phenol:chloroform:-isoamyl alcohol (24:24:1) and once with chloroform, and precipitated with 1/10 volume of 3 M NaAcetate, pH 5.2, and 2 volumes of ethanol.

Fifty µg of each RNA from the autopic tissues are DNase treated with the DNA-free kit purchased from Ambion (Austin, TX). After resuspension and spectrophotometric quantification, each sample is reverse transcribed with the TaqMan Reverse Transcription Reagents (PE Applied Biosystems, CA) according to the manufacturer's protocol. The final concentration of RNA in the reaction mix is

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200 ng/ μ l. Reverse transcription is carried out with 2.5 μ M of random hexamer primers.

TaqMan quantitative analysis. Specific primers and probe are designed according to the recommendations of PE Applied Biosystems; the probe can be labeled at the 5' end FAM (6-carboxy-fluorescein) and at the 3' end with TAMRA (6-carboxy-tetramethyl-rhodamine). Quantification experiments are performed on 10 ng of reverse transcribed RNA from each sample. Each determination is done in triplicate.

Total cDNA content is normalized with the simultaneous quantification (multiplex PCR) of the 18S ribosomal RNA using the Pre-Developed TaqMan Assay Reagents (PDAR) Control Kit (PE Applied Biosystems, CA).

The assay reaction mix is as follows: 1X final TaqMan Universal PCR Master Mix (from 2X stock) (PE Applied Biosystems, CA); 1X PDAR control – 18S RNA (from 20X stock); 300 nM forward primer; 900 nM reverse primer; 200 nM probe; 10 ng cDNA; and water to 25 μ l.

Each of the following steps are carried out once: pre PCR, 2 minutes at 50°C, and 10 minutes at 95°C. The following steps are carried out 40 times: denaturation, 15 seconds at 95°C, annealing/extension, 1 minute at 60°C.

The experiment is performed on an ABI Prism 7700 Sequence Detector (PE Applied Biosystems, CA). At the end of the run, fluorescence data acquired during PCR are processed as described in the ABI Prism 7700 user's manual in order to achieve better background subtraction as well as signal linearity with the starting target quantity.

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EXAMPLE 7*Diabetes: In vivo testing of compounds/target validation***5. 1. Glucose Production**

Over-production of glucose by the liver, due to an enhanced rate of gluconeogenesis, is the major cause of fasting hyperglycemia in diabetes. Overnight fasted normal rats or mice have elevated rates of gluconeogenesis as do streptozotocin-induced diabetic rats or mice fed ad libitum. Rats are made diabetic with a single intravenous injection of 40 mg/kg of streptozotocin while C57BL/KSJ mice are given 40-60 mg/kg ip for 5 consecutive days.

Blood glucose is measured from tail-tip blood and then compounds are administered via different routes (p.o., i.p., i.v., s.c.). Blood is collected at various times thereafter and glucose measured. Alternatively, compounds are administered for several days, then the animals are fasted overnight, blood is collected and plasma glucose measured. Compounds that inhibit glucose production will decrease plasma glucose levels compared to the vehicle-treated control group.

10 2. Insulin Sensitivity

Both ob/ob and db/db mice as well as diabetic Zucker rats are hyperglycemic, hyperinsulinemic and insulin resistant. The animals are pre-bled, their glucose levels measured, and then they are grouped so that the mean glucose level is the same for each group. Compounds are administered daily either q.d. or b.i.d. by different routes (p.o., i.p., s.c.) for 7-28 days. Blood is collected at various times and plasma glucose and insulin levels determined. Compounds that improve insulin sensitivity in these models will decrease both plasma glucose and insulin levels when compared to the vehicle-treated control group.

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3. Insulin Secretion

Compounds that enhance insulin secretion from the pancreas will increase plasma insulin levels and improve the disappearance of plasma glucose following the administration of a glucose load. When measuring insulin levels, compounds are administered by different routes (p.o., i.p., s.c. or i.v.) to overnight fasted normal rats or mice. At the appropriate time an intravenous glucose load (0.4g/kg) is given, blood is collected one minute later. Plasma insulin levels are determined. Compounds that enhance insulin secretion will increase plasma insulin levels compared to animals given only glucose. When measuring glucose disappearance, animals are bled at the appropriate time after compound administration, then given either an oral or intraperitoneal glucose load (1 g/kg), bled again after 15, 30, 60 and 90 minutes and plasma glucose levels determined. Compounds that increase insulin levels will decrease glucose levels and the area-under-the glucose curve when compared to the vehicle-treated group given only glucose.

4. Glucose Production

5 Compounds that enhance insulin secretion from the pancreas will increase plasma insulin levels and improve the disappearance of plasma glucose following the administration of a glucose load. When measuring insulin levels, compounds are administered by different routes (p.o., i.p., s.c. or i.v.) to overnight fasted normal rats or mice. At the appropriate time an intravenous glucose load (0.4g/kg) is given, blood is collected one minute later. Plasma insulin levels are determined. Compounds that enhance insulin secretion will increase plasma insulin levels compared to animals given only glucose. When measuring glucose disappearance, animals are bled at the appropriate time after compound administration, then given either an oral or intraperitoneal glucose load (1 g/kg), bled again after 15, 30, 60 and 90 minutes and plasma glucose levels determined. Compounds that increase insulin levels will decrease glucose levels and the area-under-the glucose curve when compared to the vehicle-treated group given only glucose.

10 Over-production of glucose by the liver, due to an enhanced rate of gluconeogenesis, is the major cause of fasting hyperglycemia in diabetes. Overnight fasted normal rats or mice have elevated rates of gluconeogenesis as do streptozotocin-induced diabetic rats or mice fed ad libitum. Rats are made diabetic with a single intravenous injection of 40 mg/kg of streptozotocin while C57BL/KSJ mice are given 40-60 mg/kg i.p. for 5 consecutive days.

15 Blood glucose is measured from tail-tip blood and then compounds are administered via different routes (p.o., i.p., i.v., s.c.). Blood is collected at various times thereafter and glucose measured. Alternatively, compounds are administered for several days, then the animals are fasted overnight, blood is collected and plasma glucose measured. Compounds that inhibit glucose production will decrease plasma glucose levels compared to the vehicle-treated control group.

20 Insulin Sensitivity

20 Both ob/ob and db/db mice as well as diabetic Zucker rats are hyperglycemic, hyperinsulinemic and insulin resistant. The animals are pre-bled, their glucose levels measured, and then they are grouped so that the mean glucose level is the same for each group. Compounds are administered daily either q.d. or b.i.d. by different routes (p.o., i.p., s.c.) for 7-28 days. Blood is collected at various times and plasma glucose and insulin levels determined. Compounds that improve insulin sensitivity in these models will decrease both plasma glucose and insulin levels when compared to the vehicle-treated control group.

25 Compounds that enhance insulin secretion will increase plasma insulin levels compared to animals given only glucose. When measuring glucose disappearance, animals are bled at the appropriate time after compound administration, then given either an oral or intraperitoneal glucose load (1 g/kg), bled again after 15, 30, 60, and 90 minutes and plasma glucose levels determined. Test compounds that increase insulin levels will

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decrease glucose levels and the area-under-the glucose curve when compared to the vehicle-treated group given only glucose.

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6. Insulin Secretion

Compounds that enhance insulin secretion from the pancreas will increase plasma insulin levels and improve the disappearance of plasma glucose following the administration of a glucose load. When measuring insulin levels, compounds are administered by different routes (p.o., i.p., s.c. or i.v.) to overnight fasted normal rats or mice. At the appropriate time an intravenous glucose load (0.4 g/kg) is given, blood is collected one minute later. Plasma insulin levels are determined. Compounds that enhance insulin secretion will increase plasma insulin levels compared to animals given only glucose. When measuring glucose disappearance, animals are bled at the appropriate time after compound administration, then given either an oral or intraperitoneal glucose load (1 g/kg), bled again after 15, 30, 60 and 90 minutes and plasma glucose levels determined. Compounds that increase insulin levels will decrease glucose levels and the area under the glucose curve when compared to the vehicle-treated group given only glucose.

EXAMPLE 8***In vivo testing of compounds/target validation*****1. Pain*****Acute Pain***

Acute pain is measured on a hot plate mainly in rats. Two variants of hot plate testing are used: In the classical variant animals are put on a hot surface (52 to 56°C) and the latency time is measured until the animals show nociceptive behavior, such as stepping or foot licking. The other variant is an increasing temperature hot plate where the experimental animals are put on a surface of neutral temperature. Subsequently this surface is slowly but constantly heated until the animals begin to lick a hind paw. The temperature which is reached when hind paw licking begins is a measure for pain threshold.

Compounds are tested against a vehicle treated control group. Substance 5 application is performed at different time points via different application routes (i.v., i.p., p.o., i.t., i.c.v., s.c., intradermal, transdermal) prior to pain testing.

Persistent Pain

10 Persistent pain is measured with the formalin or capsaicin test, mainly in rats. A solution of 1 to 5% formalin or 10 to 100 µg capsaicin is injected into one hind paw of the experimental animal. After formalin or capsaicin application the animals show nociceptive reactions like flinching, licking and biting of the affected paw. The number of nociceptive reactions within a time frame of up to 90 minutes is a measure for intensity of pain.

Compounds are tested against a vehicle treated control group. Substance 15 application is performed at different time points via different application routes (i.v., i.p., p.o., i.t., i.c.v., s.c., intradermal, transdermal) prior to formalin or capsaicin administration.

Neuropathic Pain

25 Neuropathic pain is induced by different variants of unilateral sciatic nerve injury mainly in rats. The operation is performed under anesthesia. The first variant of sciatic nerve injury is produced by placing loosely constrictive ligatures around the common sciatic nerve. The second variant is the tight ligation of about the half of the diameter of the common sciatic nerve. In the next variant, a group of models is used in which tight ligations or transections are made of either the L5 and L6 spinal nerves, or the L1% spinal nerve only.

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The fourth variant involves an axotomy of two of the three terminal branches of the sciatic nerve (tibial and common peroneal nerves) leaving the remaining sural nerve intact whereas the last variant comprises the axotomy of only the tibial branch leaving the sural and common nerves uninjured. Control animals are treated with a sham operation.

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Postoperatively, the nerve injured animals develop a chronic mechanical allodynia, cold allodynia, as well as a thermal hyperalgesia. Mechanical allodynia is measured by means of a pressure transducer (electronic von Frey Anesthesiometer, IITC Inc.-Life Science Instruments, Woodland Hills, SA, USA). Thermal hyperalgesia is measured by means of a radiant heat source (Plantar Test, Ugo Basile, Comerio, Italy, Paw thermal stimulator, G. Ozaki, University of California, USA). For edema measurement two methods are being used. In the first method, the animals are sacrificed and the affected hindpaws sectioned and weighed. The second method comprises differences in paw volume by measuring water displacement in a plethysmometer (Ugo Basile, Comerio, Italy).

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Thermal hyperalgesia is measured by means of a radiant heat source (Plantar Test, Ugo Basile, Comerio, Italy), or by means of a cold plate of 5 to 10°C where the nociceptive reactions of the affected hind paw are counted as a measure of pain intensity. A further test for cold induced pain is the counting of nociceptive reactions, or duration of nociceptive responses after plantar administration of acetone to the affected hind limb. Chronic pain in general is assessed by registering the circadian rhythms in activity (Sutrio and Arndt, Universitä zu Köln, Cologne, Germany), and by scoring differences in gait (foot print patterns; FOOTPRINTS program, Klapdor et al, 1997. A low cost method to analyze footprint patterns. *J. Neurosci. Methods* 75, 49-54).

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Compounds are tested against sham operated and vehicle treated control groups. Substance application is performed at different time points via different application routes (i.v., i.p., p.o., it., i.c.v., s.c., intradermal, transdermal) prior to pain testing.

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Inflammatory Pain

Inflammatory pain is induced mainly in rats by injection of 0.75 mg carrageenan or complete Freund's adjuvant into one hind paw. The animals

develop an edema with mechanical allodynia as well as thermal hyperalgesia. Mechanical allodynia is measured by means of a pressure transducer (electronic von Frey Anesthesiometer, IITC Inc.-Life Science Instruments, Woodland Hills, SA, USA). Thermal hyperalgesia is measured by means of a radiant heat source (Plantar Test, Ugo Basile, Comerio, Italy, Paw thermal stimulator, G. Ozaki, University of California, USA). For edema measurement two methods are being used. In the first method, the animals are sacrificed and the affected hindpaws sectioned and weighed. The second method comprises differences in paw volume by measuring water displacement in a plethysmometer (Ugo Basile, Comerio, Italy).

Compounds are tested against uninflamed as well as vehicle treated control groups. Substance application is performed at different time points via different application routes (i.v., i.p., p.o., it., i.c.v., s.c., intradermal, transdermal) prior to pain testing.

Diabetic Neuropathic Pain

Rats treated with a single intraperitoneal injection of 50 to 80 mg/kg streptozotocin develop a profound hyperglycemia and mechanical allodynia within 1 to 3 weeks. Mechanical allodynia is measured by means of a pressure transducer (electronic von Frey Anesthesiometer, IITC Inc.-Life Science Instruments, Woodland Hills, SA, USA).

Compounds are tested against diabetic and non-diabetic vehicle treated control groups. Substance application is performed at different time points via different application routes (i.v., i.p., p.o., it., i.c.v., s.c., intradermal, transdermal) prior to pain testing.

Inflammatory pain is induced mainly in rats by injection of 0.75 mg carrageenan or complete Freund's adjuvant into one hind paw. The animals

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2. Parkinson's disease

6-Hydroxydopamine (6-OH-DA) Lesion

5 Degeneration of the dopaminergic nigrostriatal and striatopallidal pathways is the central pathological event in Parkinson's disease. This disorder has been mimicked experimentally in rats using single/sequential unilateral stereotoxic injections of 6-OH-DA into the medium forebrain bundle (MFB).

10 Male Wistar rats (Harlan Winkelmann, Germany), weighing 200±250 g at the beginning of the experiment, are used. The rats are maintained in a temperature- and humidity-controlled environment under a 12 h light/dark cycle with free access to food and water when not in experimental sessions. The following in vivo protocols are approved by the governmental authorities.

15 All efforts are made to minimize animal suffering, to reduce the number of animals used, and to utilize alternatives to in vivo techniques.

5 Forelimb ataxia is assessed three weeks following lesion placement using a modified stepping test protocol. In brief, the animals are held by the experimenter with one hand fixing the hindlimbs and slightly raising the hind part above the surface. One paw is touching the table, and is then moved slowly sideways (5 s for 1 m), first in the forehand and then in the backhand direction. The number of adjusting steps is counted for both paws in the backhand and forehand direction of movement. The sequence of testing is right paw forehand and backhand adjusting stepping, followed by left paw forehand and backhand directions. The test is repeated three times on three consecutive days, after an initial training period of three days prior to the first testing. Forehand adjusted stepping reveals no consistent differences between lesioned and healthy control animals. Analysis is therefore restricted to backhand adjusted stepping.

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Animals are administered pargyline on the day of surgery (Sigma, St. Louis, MO, USA; 50 mg/kg i.p.) in order to inhibit metabolism of 6-OHDA by monoamine oxidase and desmethylimipramine HCl (Sigma; 25 mg/kg i.p.) in order to prevent uptake of 6-OHDA by noradrenergic terminals. Thirty minutes later the rats are anesthetized with sodium pentobarbital (50 mg/kg) and placed in a stereotoxic frame. In order to lesion the DA nigrostriatal pathway 4 µl of 0.01% ascorbic acid-saline containing 8 µg of 6-OHDA HBr (Sigma) are injected into the left medial fore-brain bundle at a rate of 1 µl/min (2.4 mm anterior, 1.49 mm lateral, -2.7 mm ventral to Bregma and the skull surface). The needle is left in place an additional 5 min to allow diffusion to occur.

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20 Balance adjustments following postural challenge are also measured during the stepping test sessions. The rats are held in the same position as described in the stepping test and, instead of being moved sideways, tilted by the experimenter towards the side of the paw touching the table. This maneuver results in loss of balance and the ability of the rats to regain balance by forelimb movements is scored on a scale ranging from 0 to 3. Score 0 is given for a normal forelimb placement. When the forelimb movement is delayed but recovery of postural balance detected, score 1 is given. Score 2 represents a clear, yet insufficient, forelimb reaction, as evidenced by muscle contraction, but lack of success in recovering balance, and score 3 is given for no reaction of movement. The test is repeated three times a day on each side for three consecutive days after an initial training period of three days prior to the first testing.

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Stepping Test

Balance Test

Balance adjustments following postural challenge are also measured during the stepping test sessions. The rats are held in the same position as described in the stepping test and, instead of being moved sideways, tilted by the experimenter towards the side of the paw touching the table. This maneuver results in loss of balance and the ability of the rats to regain balance by forelimb movements is scored on a scale ranging from 0 to 3. Score 0 is given for a normal forelimb placement. When the forelimb movement is delayed but

recovers of postural balance detected, score 1 is given. Score 2 represents a clear, yet insufficient, forelimb reaction, as evidenced by muscle contraction, but lack of success in recovering balance, and score 3 is given for no reaction of movement. The test is repeated three times a day on each side for three consecutive days after an initial training period of three days prior to the first testing.

Staircase Test (Paw Beaching)

A modified version of the staircase test is used for evaluation of paw reaching behavior three weeks following primary and secondary lesion placement.

Plexiglass test boxes with a central platform and a removable staircase on each side are used. The apparatus is designed such that only the paw on the same side at each staircase can be used, thus providing a measure of independent forelimb use. For each test the animals are left in the test boxes for 15 min. The double staircase is filled with 7 x 3 chow pellets (Precision food pellets, formula: P, purified rodent diet, size 45 mg; Sandown Scientific) on each side. After each test the number of pellets eaten (successfully retrieved pellets) and the number of pellets taken (touched but dropped) for each paw and the success rate (pellets eaten/pellets taken) are counted separately. After three days of food deprivation (12 g per animal per day) the animals are tested for 11 days. Full analysis is conducted only for the last five days.

MPTP treatment

The neurotoxin 1-methyl-4-phenyl-1,2,3,6-tetrahydro-pyridine (MPTP) causes degeneration of mesencephalic dopaminergic (DAergic) neurons in rodents, non-human primates, and humans and, in so doing, reproduces many of the symptoms of Parkinson's disease. MPTP leads to a marked decrease in the levels of dopamine and its metabolites, and in the number of dopaminergic terminals in the striatum as well as severe loss of the tyrosine hydroxylase (TH)-immunoreactive cell bodies in the substantia nigra, pars compacta.

lesion 7–10 days later. Successive MPTP injections are administered on days 1, 2 and 3. Animals receive application of 4 mg/kg MPTP hydrochloride (Sigma) in saline once daily. All injections are intraperitoneal (i.p.) and the MPTP stock solution is frozen between injections. Animals are decapitated on day 11.

Immunohistology

At the completion of behavioral experiments, all animals are anaesthetized with 3 ml thiopental (1 g/40 ml i.p., Tyrol Pharma). The mice are perfused transcardially with 0.01 M PBS (pH 7.4) for 2 min, followed by 4% paraformaldehyde (Merck) in PBS for 15 min. The brains are removed and placed in 4% paraformaldehyde for 24 h at 4°C. For dehydration they are then transferred to a 20% sucrose (Merck) solution in 0.1 M PBS at 4°C until they sink. The brains are frozen in methylbutan at -20°C for 2 min and stored at -70°C. Using a sledge microtome (mod. 3800-Frigocut, Leica), 25 µm sections are taken from the genu of the corpus callosum (AP 1.7 mm) to the hippocampus (AP 21.8 mm) and from AP 24.16 to AP 26.72. Forty-six sections are cut and stored in assorters in 0.25 M Tris buffer (pH 7.4) for

A series of sections is processed for free-floating tyrosine hydroxylase (TH) immunohistochemistry. Following three rinses in 0.1 M PBS, endogenous peroxidase activity is quenched for 10 min in 0.3% $H_2O_2 \pm$ PBS. After rinsing in PBS, sections are preincubated in 10% normal bovine serum (Sigma) for 5 min as blocking agent and transferred to either primary anti-rat TH rabbit antiserum (dilution 1:2000).

In order to obtain severe and long-lasting lesions, and to reduce mortality, animals receive single injections of MPTP, and are then tested for severity of

Following overnight incubation at room temperature, sections for TH immunoreactivity are rinsed in PBS (2×10 min) and incubated in biotinylated anti-rabbit immunoglobulin G raised in goat (dilution 1:200) (Vector) for 90

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min, rinsed repeatedly and transferred to Vectastain ABC (Vector) solution for 1 h. 3,3' -Diaminobenzidine tetrahydrochloride (DAB; Sigma) in 0.1 M PBS, supplemented with 0.005% H₂O₂, serves as chromogen in the subsequent visualization reaction. Sections are mounted on to gelatin-coated slides, left to dry overnight, counter-stained with hematoxylin dehydrated in ascending alcohol concentrations and cleared in butylacetate. Coverslips are mounted on entellan.

Rotarod Test

We use a modification of the procedure described by Rozas and Labandeira-Garcia (1997), with a CR-1 Rotamex system (Columbus Instruments, Columbus, OH) comprising an IBM-compatible personal computer, a CIO-24 data acquisition card, a control unit, and a four-lane rotarod unit. The rotarod unit consists of a rotating spindle (diameter 7.3 cm) and individual compartments for each mouse. The system software allows preprogramming of session protocols with varying rotational speeds (0–80 rpm). Infrared beams are used to detect when a mouse has fallen onto the base grid beneath the rotarod. The system logs the fall as the end of the experiment for that mouse, and the total time on the rotarod, as well as the time of the fall and all the set-up parameters, are recorded. The system also allows a weak current to be passed through the base grid, to aid training.

3. Dementia***The object recognition task***

The object recognition task has been designed to assess the effects of experimental manipulations on the cognitive performance of rodents. A rat is placed in an open field, in which two identical objects are present. The rats inspects both objects during the first trial of the object recognition task. In a second

trial, after a retention interval of for example 24 hours, one of the two objects used in the first trial, the 'familiar' object, and a novel object are placed in the open field. The inspection time at each of the objects is registered. The basic measures in the OR task is the time spent by a rat exploring the two object the second trial. Good retention is reflected by higher exploration times towards the novel than the 'familiar' object.

Administration of the putative cognition enhancer prior to the first trial predominantly allows assessment of the effects on acquisition, and eventually on consolidation processes. Administration of the testing compound after the first trial allows to assess the effects on consolidation processes, whereas administration before the second trial allows to measure effects on retrieval processes.

The passive avoidance task

The passive avoidance task assesses memory performance in rats and mice. The inhibitory avoidance apparatus consists of a two-compartment box with a light compartment and a dark compartment. The two compartments are separated by a guillotine door that can be operated by the experimenter. A threshold of 2 cm separates the two compartments when the guillotine door is raised. When the door is open, the illumination in the dark compartment is about 2 lux. The light intensity is about 500 lux at the center of the floor of the light compartment.

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Two habituation sessions, one shock session, and a retention session are given, separated by inter-session intervals of 24 hours. In the habituation sessions and the retention session the rat is allowed to explore the apparatus for 300 sec. The rat is placed in the light compartment, facing the wall opposite to the guillotine door. After an accommodation period of 15 sec, the guillotine door is opened so that all parts of the apparatus can be visited

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freely. Rats normally avoid brightly lit areas and will enter the dark compartment within a few seconds.

In the shock session the guillotine door between the compartments is lowered as soon as the rat has entered the dark compartment with its four paws, and a scrambled 1 mA footshock is administered for 2 sec. The rat is removed from the apparatus and put back into its home cage. The procedure during the retention session is identical to that of the habituation sessions.

The step-through latency, that is the first latency of entering the dark compartment (in sec.) during the retention session is an index of the memory performance of the animal; the longer the latency to enter the dark compartment, the better the retention is. A testing compound is given half an hour before the shock session, together with 1 mg*kg⁻¹ scopolamine. Scopolamine impairs the memory performance during the retention session 24 hours later. If the test compound increases the enter latency compared with the scopolamine-treated controls, is likely to possess cognition enhancing potential.

The Morris water escape task

The Morris water escape task measures spatial orientation learning in rodents. It is a test system that has extensively been used to investigate the effects of putative therapeutics on the cognitive functions of rats and mice. The performance of an animal is assessed in a circular water tank with an escape platform that is submerged about 1 cm below the surface of the water. The escape platform is not visible for an animal swimming in the water tank. Abundant extra-maze cues are provided by the furniture in the room, including desks, computer equipment, a second water tank, the presence of the experimenter, and by a radio on a shelf that is playing softly.

The animals receive four trials during five daily acquisition sessions. A trial is started by placing an animal into the pool, facing the wall of the tank. Each of four starting positions in the quadrants north, east, south, and west is used once in a series of four trials; their order is randomized. The escape platform is always in the same position. A trial is terminated as soon as the animal had climbed onto the escape platform or when 90 seconds have elapsed, whichever event occurs first. The animal is allowed to stay on the platform for 30 seconds. Then it is taken from the platform and the next trial is started. If an animal did not find the platform within 90 seconds it is put on the platform by the experimenter and is allowed to stay there for 30 seconds. After the fourth trial of the fifth daily session, an additional trial is given as a probe trial: the platform is removed, and the time the animal spends in the four quadrants is measured for 30 or 60 seconds. In the probe trial, all animals start from the same start position, opposite to the quadrant where the escape platform had been positioned during acquisition.

Four different measures are taken to evaluate the performance of an animal during acquisition training: escape latency, traveled distance, distance to platform, and swimming speed. The following measures are evaluated for the probe trial: time (s) in quadrants and traveled distance (cm) in the four quadrants. The probe trial provides additional information about how well an animal learned the position of the escape platform. If an animal spends more time and swims a longer distance in the quadrant where the platform had been positioned during the acquisition sessions than in any other quadrant, one concludes that the platform position has been learned well.

In order to assess the effects of putative cognition enhancing compounds, rats or mice with specific brain lesions which impair cognitive functions, or animals treated with compounds such as scopolamine or MK-801, which interfere with normal learning, or aged animals which suffer from cognitive deficits, are used.

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The T-maze spontaneous alternation task

The T-maze spontaneous alternation task (TeMCAT) assesses the spatial memory performance in mice. The start arm and the two goal arms of the T-maze are provided with guillotine doors which can be operated manually by the experimenter. A mouse is put into the start arm at the beginning of training. The guillotine door is closed. In the first trial, the 'forced trial', either the left or right goal arm is blocked by lowering the guillotine door.

After the mouse has been released from the start arm, it will negotiate the maze, eventually enter the open goal arm, and return to the start position, where it will be confined for 5 seconds, by lowering the guillotine door. Then, the animal can choose freely between the left and right goal arm (all guillotine-doors opened) during 14 'free choice' trials. As soon as the mouse has entered one goal arm, the other one is closed. The mouse eventually returns to the start arm and is free to visit whichever goal arm it wants after having been confined to the start arm for 5 seconds. After completion of 14 free choice trials in one session, the animal is removed from the maze. During training, the animal is never handled.

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The percent alternations out of 14 trials is calculated. This percentage and the total time needed to complete the first forced trial and the subsequent 14 free choice trials (in s) is analyzed. Cognitive deficits are usually induced by an injection of scopolamine, 30 min before the start of the training session. Scopolamine reduced the per-cent alternations to chance level, or below. A cognition enhancer, which is always administered before the training session, will at least partially, antagonize the scopolamine-induced reduction in the spontaneous alternation rate.

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The addition of the test oligonucleotide for seven days results in significantly reduced expression of human elongase HSEL01-like protein as determined by Western blotting. This effect is not observed with the control oligonucleotide. After 3 to 7 days, the number of cells in the cultures is counted using an automatic cell counter. The number of cells in cultures treated with the test oligonucleotide (expressed as 100%) is compared with the number of cells in cultures treated with the control oligonucleotide. The number of cells in cultures treated with the test oligonucleotide is not more than 30% of control, indicating that the inhibition of human elongase HSEL01-like protein has an anti-proliferative effect on cancer cells.

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EXAMPLE 9

Proliferation inhibition assay: Antisense oligonucleotides suppress the growth of cancer cell lines

The cell line used for testing is the human colon cancer cell line HCT116. Cells are cultured in RPMI-1640 with 10-15% fetal calf serum at a concentration of 10,000 cells per milliliter in a volume of 0.5 ml and kept at 37°C in a 95% air/5%CO₂ atmosphere.

Phosphothioate oligoribonucleotides are synthesized on an Applied Biosystems Model 380B DNA synthesizer using phosphoroamidite chemistry. A sequence of 24 bases complementary to the nucleotides at position 1 to 24 of SEQ ID NO: 1 is used as the test oligonucleotide. As a control, another (random) sequence is used: 5'-TCA ACT GAC TAG ATG TAC ATG GAC-3'. Following assembly and deprotection, oligonucleotides are ethanol-precipitated twice, dried, and suspended in phosphate buffered saline at the desired concentration. Purity of the oligonucleotides is tested by capillary gel electrophoresis and ion exchange HPLC. The purified oligonucleotides are added to the culture medium at a concentration of 10 μM once per day for seven days.

The addition of the test oligonucleotide for seven days results in significantly reduced expression of human elongase HSEL01-like protein as determined by Western blotting. This effect is not observed with the control oligonucleotide. After 3 to 7 days, the number of cells in the cultures is counted using an automatic cell counter. The number of cells in cultures treated with the test oligonucleotide (expressed as 100%) is compared with the number of cells in cultures treated with the control oligonucleotide. The number of cells in cultures treated with the test oligonucleotide is not more than 30% of control, indicating that the inhibition of human elongase HSEL01-like protein has an anti-proliferative effect on cancer cells.

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EXAMPLE 10

In vivo testing of compounds/large validation

Rank Sum test after the variance between groups is compared by an F-test, with significance at $p < 0.05$ as compared to the vehicle control group.

Anti-Mechanistic Annotations

This non-tumor assay measures the ability of a compound to reduce either the endogenous level of a circulating hormone or the level of hormone produced in response to a biologic stimulus. Rodents are administered test compound (p.o., i.p., i.v., i.m., or s.c.). At a predetermined time after administration of test compound, blood plasma is collected. Plasma is assayed for levels of the hormone of interest. If the normal circulating levels of the hormone are too low and/or variable to provide consistent results, the level of the hormone may be elevated by a pre-treatment with a biologic stimulus (i.e., LHRH may be injected i.m. into mice at a dosage of 30 ng/mouse to induce a burst of testosterone synthesis). The timing of plasma collection would be adjusted to coincide with the peak of the induced hormone response. Compound effects are compared to a vehicle-treated control group. An F-test is performed to determine if the variance is equal or unequal followed by a Student's t-test. Significance is p value \leq 0.05 compared to the vehicle control group.

1. Acute Mechanistic Assays

1.1. Reduction in Mitogenic Plasma Hormone Levels

This is another non-tumor assay that measures the ability of a compound to reduce the mass of a hormone dependent tissue (i.e., seminal vesicles in males and uterus in females). Rodents are administered test compound (p.o., i.p., i.v., i.m., or s.c.) according to a predetermined schedule and for a predetermined duration (i.e., 1 week). At termination of the study, animals are weighed, the target organ is excised, any fluid is expressed, and the weight of the organ is recorded. Blood plasma may also be collected. Plasma may be assayed for levels of a hormone of interest or for levels of test agent. Organ weights may be directly compared or they may be normalized for the body weight of the animal. Compound effects are compared to a vehicle-treated control group. An F-test is performed to determine if the variance is equal or unequal followed by a Student's t-test. Significance is p value \leq 0.05 compared to the vehicle control group.

2. Subacute Functional *In Vivo* Assays

3.1 Reduction in Mass of Hormone Dependent Tissues

2. Subacute Functional *In Vivo* Assays

3.1 Reduction in Mass of Hormone Dependent Tissues

1.2. Hollow Fiber Mechanism of Action Assay

Hollow fibers are prepared with desired cell line(s) and implanted intraperitoneally and/or subcutaneously in rodents. Compounds are administered p.o., i.p., i.v., i.m., or s.c. Fibers are harvested in accordance with specific readout assay protocol, these may include assays for gene expression (bDNA, PCR, or Taqman), or a specific biochemical activity (i.e. cAMP levels). Results are analyzed by Student's t-test or

1.2. Hollow Fiber Mechanism of Action Assay

proliferation is determined by measuring a marker of cell number (i.e., MTT or LDH). The cell number and change in cell number from the starting inoculum are analyzed by Student's t-test or Rank Sum test after the variance between groups is compared by an F-test, with significance at $p \leq 0.05$ as compared to the vehicle control group.

Hollow fibers are prepared with desired cell line(s) and implanted intraperitoneally and/or subcutaneously in rodents. Compounds are administered p.o., i.p., i.v., i.m., or s.c. Fibers are harvested in accordance with specific readout assay protocol. Cell proliferation is determined by measuring a marker of cell number (i.e., MTT or LDH). The cell number and change in cell number from the starting inoculum are analyzed by Student's t-test or Rank Sum test after the variance between groups is compared by an F-test, with significance at $p \leq 0.05$ as compared to the vehicle control group.

and/or subcutaneously in rodents. Compounds are administered p.o., i.p., i.v., i.m., or

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recorded 2-3 times weekly. Mean tumor weights of all groups over days post inoculation are graphed for comparison. An F-test is preformed to determine if the variance is equal or unequal followed by a Student's t-test to compare tumor sizes in the treated and control groups at the end of treatment. Significance is $p \leq 0.05$ as compared to the control group. Tumor measurements may be recorded after dosing has stopped to monitor tumor growth delay. Tumor growth delays are expressed as the difference in the median time for the treated and control groups to attain a predetermined size divided by the median time for the control group to attain that size. Growth delays are compared by generating Kaplan-Meier curves from the times for individual tumors to attain the evaluation size. Significance is $p \leq 0.05$ as compared to the vehicle control group.

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recorded 2-3 times weekly. Mean tumor weights of all groups over days post inoculation are graphed for comparison. An F-test is preformed to determine if the variance is equal or unequal followed by a Student's t-test to compare tumor sizes in the treated and control groups at the end of treatment. Significance is $p \leq 0.05$ as compared to the control group. Tumor measurements may be recorded after dosing has stopped to monitor tumor growth delay. Tumor growth delays are expressed as the difference in the median time for the treated and control groups to attain a predetermined size divided by the median time for the control group to attain that size. Growth delays are compared by generating Kaplan-Meier curves from the times for individual tumors to attain the evaluation size. Significance is $p \leq 0.05$ as compared to the vehicle control group.

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3.3. Orthotopic Disease Models

3.3.1. Mammary Fat Pad Assay

Tumor cells or fragments, of mammary adenocarcinoma origin, are implanted directly into a surgically exposed and reflected mammary fat pad in rodents. The fat pad is placed back in its original position and the surgical site is closed. Hormones may also be administered to the rodents to support the growth of the tumors. Compounds are administered p.o., i.p., i.v., i.m., or s.c. according to a predetermined schedule. Body weights are measured and recorded 2-3 times weekly. Tumor and body weights are measured and recorded 2-3 times weekly. Mean tumor weights of all groups over days post inoculation are graphed for comparison. An F-test is preformed to determine if the variance is equal or unequal followed by a Student's t-test to compare tumor sizes in the treated and control groups at the end of treatment. Significance is $p \leq 0.05$ as compared to the control group.

Tumor measurements may be recorded after dosing has stopped to monitor tumor growth delay. Tumor growth delays are expressed as the difference in the median time for the treated and control groups to attain a predetermined size divided by the

3.3.2. Intraprostatic Assay

Tumor cells or fragments, of prostatic adenocarcinoma origin, are implanted directly into a surgically exposed dorsal lobe of the prostate in rodents. The prostate is externalized through an abdominal incision so that the tumor can be implanted specifically in the dorsal lobe while verifying that the implant does not enter the seminal vesicles. The successfully inoculated prostate is replaced in the abdomen and the incisions through the abdomen and skin are closed. Hormones may also be administered to the rodents to support the growth of the tumors. Compounds are administered p.o., i.p., i.v., i.m., or s.c. according to a predetermined schedule. Body weights are measured and recorded 2-3 times weekly. At a predetermined time, the experiment is terminated and the animal is dissected. The size of the primary tumor is measured in three dimensions using either a caliper or an ocular micrometer attached to a dissecting scope. An F-test is performed to determine if the variance is equal or unequal followed by a Student's t-test to compare tumor sizes in the treated and control groups at the end of treatment. Significance is $p \leq 0.05$ as compared to the control group. This model provides an opportunity to increase the rate of spontaneous metastasis of this type of tumor. Metastasis can be assessed at termination of the study by counting the number of visible foci per target organ (i.e., the lungs), or measuring the target organ weight (i.e., the regional lymph nodes). The means of these endpoints are compared by Student's t-test after conducting an F-test,

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with significance determined at $p \leq 0.05$ compared to the control group in the experiment.

3.3.3. *Intrabronchial Assay*

Tumor cells of pulmonary origin may be implanted intrabronchially by making an incision through the skin and exposing the trachea. The trachea is pierced with the beveled end of a 25 gauge needle and the tumor cells are inoculated into the main bronchus using a flat-ended 27 gauge needle with a 90° bend. Compounds are administered p.o., i.p., i.v., i.m., or s.c. according to a predetermined schedule. Body weights are measured and recorded 2-3 times weekly. At a predetermined time, the experiment is terminated and the animal is dissected. The size of the primary tumor is measured in three dimensions using either a caliper or an ocular micrometer attached to a dissecting scope. An F-test is preformed to determine if the variance is equal or unequal followed by a Student's t-test to compare tumor sizes in the treated and control groups at the end of treatment. Significance is $p \leq 0.05$ as compared to the control group. This model provides an opportunity to increase the rate of spontaneous metastasis of this type of tumor.

5 Metastasis can be assessed at termination of the study by counting the number of visible foci per target organ (i.e., the liver), or measuring the target organ weight. The means of these endpoints are compared by Student's t-test after conducting an F-test, with significance determined at $p \leq 0.05$ compared to the control group in the experiment.

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Metastasis can be assessed at termination of the study by counting the number of visible foci per target organ (i.e., the lung or liver). These primary tumors are then excised. Compounds are administered p.o., i.p., i.v., i.m., or s.c. according to a predetermined schedule which may include the period leading up to the excision of the primary tumor to evaluate therapies directed at inhibiting the early stages of tumor metastasis. Observations of morbidity and/or mortality are recorded daily. Body weights are measured and recorded twice weekly. Potential endpoints include survival time, numbers of visible foci per target organ, or target organ weight. When survival time is used as the endpoint the other values are not determined. Survival

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data is used to generate Kaplan-Meier curves. Significance is $p \leq 0.05$ by a log-rank test compared to the control group in the experiment. The mean number of visible tumor foci, as determined under a dissecting microscope, and the mean target organ weights are compared by Student's t-test after conducting an F-test, with significance determined at $p \leq 0.05$ compared to the control group in the experiment for both of these endpoints.

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3.3.4. *Intracecal Assay*

Tumor cells of gastrointestinal origin may be implanted intracecally by making an abdominal incision through the skin and externalizing the intestine. Tumor cells are inoculated into the cecal wall without penetrating the lumen of the intestine using a 27 or 30 gauge needle. Compounds are administered p.o., i.p., i.v., i.m., or s.c. according to a predetermined schedule. Body weights are measured and recorded 2-3 times weekly. At a predetermined time, the experiment is terminated and the animal

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is dissected. The size of the primary tumor is measured in three dimensions using either a caliper or an ocular micrometer attached to a dissecting scope. An F-test is preformed to determine if the variance is equal or unequal followed by a Student's t-test to compare tumor sizes in the treated and control groups at the end of treatment. Significance is $p \leq 0.05$ as compared to the control group. This model provides an opportunity to increase the rate of spontaneous metastasis of this type of tumor.

5 Metastasis can be assessed at termination of the study by counting the number of visible foci per target organ (i.e., the liver), or measuring the target organ weight. The means of these endpoints are compared by Student's t-test after conducting an F-test, with significance determined at $p \leq 0.05$ compared to the control group in the experiment.

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4. Secondary (Metastatic) Antitumor Efficacy

4.1. *Spontaneous Metastasis*

Tumor cells are inoculated s.c. and the tumors allowed to grow to a predetermined range for spontaneous metastasis studies to the lung or liver. These primary tumors are then excised. Compounds are administered p.o., i.p., i.v., i.m., or s.c. according to a predetermined schedule which may include the period leading up to the excision of the primary tumor to evaluate therapies directed at inhibiting the early stages of tumor metastasis. Observations of morbidity and/or mortality are recorded daily. Body weights are measured and recorded twice weekly. Potential endpoints include survival time, numbers of visible foci per target organ, or target organ weight. When survival time is used as the endpoint the other values are not determined. Survival

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data is used to generate Kaplan-Meier curves. Significance is $p \leq 0.05$ by a log-rank test compared to the control group in the experiment. The mean number of visible tumor foci, as determined under a dissecting microscope, and the mean target organ weights are compared by Student's t-test after conducting an F-test, with significance determined at $p \leq 0.05$ compared to the control group in the experiment for both of these endpoints.

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4.2. Forced Metastasis

Tumor cells are injected into the tail vein, portal vein, or the left ventricle of the heart in experimental (forced) lung, liver, and bone metastasis studies, respectively. Compounds are administered p.o., i.p., i.v., i.m., or s.c. according to a predetermined schedule. Observations of morbidity and/or mortality are recorded daily. Body weights are measured and recorded twice weekly. Potential endpoints include survival time, numbers of visible foci per target organ, or target organ weight. When survival time is used as the endpoint the other values are not determined. Survival data is used to generate Kaplan-Meier curves. Significance is $p \leq 0.05$ by a log-rank test compared to the control group in the experiment. The mean number of visible tumor foci, as determined under a dissecting microscope, and the mean target organ weights are compared by Student's t-test after conducting an F-test, with significance at $p \leq 0.05$ compared to the vehicle control group in the experiment for both endpoints.

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CLAIMS

1. An isolated polynucleotide being selected from the group consisting of:

5 a) a polynucleotide encoding a elongase HSEL01-like protein polypeptide comprising an amino acid sequence selected form the group consisting of:

10 a) amino acid sequences which are at least about 50% identical to the amino acid sequence shown in SEQ ID NO: 2; and
the amino acid sequence shown in SEQ ID NO: 2.

b) a polynucleotide comprising the sequence of SEQ ID NOS: 1, 5 or 6;

15 c) a polynucleotide which hybridizes under stringent conditions to a polynucleotide specified in (a) and (b) and encodes a elongase HSEL01-like protein polypeptide;

d) a polynucleotide the sequence of which deviates from the polynucleotide sequences specified in (a) to (c) due to the degeneration of the genetic code and encodes a elongase HSEL01-like protein polypeptide; and

e) a polynucleotide which represents a fragment, derivative or allelic variation of a polynucleotide sequence specified in (a) to (d) and encodes a elongase HSEL01-like protein polypeptide.

2. An expression vector containing any polynucleotide of claim 1.

30 3. A host cell containing the expression vector of claim 2.

4. A substantially purified elongase HSEL01-like protein polypeptide encoded by a polynucleotide of claim 1.

5. A method for producing a elongase HSEL01-like protein polypeptide, wherein the method comprises the following steps:

- a) culturing the host cell of claim 3 under conditions suitable for the expression of the elongase HSEL01-like protein polypeptide; and
- b) recovering the elongase HSEL01-like protein polypeptide from the host cell culture.

10 6. A method for detection of a polynucleotide encoding a elongase HSEL01-like protein polypeptide in a biological sample comprising the following steps:

- a) hybridizing any polynucleotide of claim 1 to a nucleic acid material of a biological sample, thereby forming a hybridization complex; and
- b) detecting said hybridization complex.

15 7. The method of claim 6, wherein before hybridization, the nucleic acid material of the biological sample is amplified.

20 8. A method for the detection of a polynucleotide of claim 1 or a elongase HSEL01-like protein polypeptide of claim 4 comprising the steps of:

- contacting a biological sample with a reagent which specifically interacts with the polynucleotide or the elongase HSEL01-like protein polypeptide.

25 9. A diagnostic kit for conducting the method of any one of claims 6 to 8.

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10. A method of screening for agents which decrease the activity of a elongase HSELO1-like protein, comprising the steps of:

5 contacting a test compound with any elongase HSELO1-like protein polypeptide encoded by any polynucleotide of claim 1;
 detecting binding of the test compound to the elongase HSELO1-like protein polypeptide, wherein a test compound which binds to the polypeptide is identified as a potential therapeutic agent for decreasing the activity of a elongase HSELO1-like protein.

10 11. A method of screening for agents which regulate the activity of a elongase HSELO1-like protein, comprising the steps of:

15 contacting a test compound with a elongase HSELO1-like protein polypeptide encoded by any polynucleotide of claim 1; and
 detecting a elongase HSELO1-like protein activity of the polypeptide, wherein a test compound which increases the elongase HSELO1-like protein activity is identified as a potential therapeutic agent for increasing the activity of the elongase HSELO1-like protein, and wherein a test compound which decreases the elongase HSELO1-like protein activity of the polypeptide is identified as a potential therapeutic agent for decreasing the activity of the elongase HSELO1-like protein.

20 12. A method of screening for agents which decrease the activity of a elongase HSELO1-like protein, comprising the steps of:

25 contacting a test compound with any polynucleotide of claim 1 and detecting binding of the test compound to the polynucleotide, wherein a test compound

which binds to the polynucleotide is identified as a potential therapeutic agent for decreasing the activity of elongase HSELO1-like protein.

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which binds to the polynucleotide is identified as a potential therapeutic agent for decreasing the activity of elongase HSELO1-like protein.

5 13. A method of reducing the activity of elongase HSELO1-like protein, comprising the steps of:

10 contacting a cell with a reagent which specifically binds to any polynucleotide of claim 1 or any elongase HSELO1-like protein polypeptide of claim 4, whereby the activity of elongase HSELO1-like protein is reduced.

15 14. A reagent that modulates the activity of a elongase HSELO1-like protein polypeptide or a polynucleotide wherein said reagent is identified by the method of any of the claim 10 to 12.

20 15. A pharmaceutical composition, comprising:
 the expression vector of claim 2 or the reagent of claim 14 and a pharmaceutically acceptable carrier.

25 16. Use of the expression vector of claim 2 or the reagent of claim 14 in the preparation of a medicament for modulating the activity of a elongase HSELO1-like protein in a disease.

30 17. Use of claim 16 wherein the disease is cancer, diabetes, CNS disorder, asthma, metabolic disease or COPD.

18. A cDNA encoding a polypeptide comprising the amino acid sequence shown in SEQ ID NC: 2.

35 19. The cDNA of claim 18 which comprises SEQ ID NOS: 1, 5 or 6.

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20. The cDNA of claim 18 which consists of SEQ ID NOS: 1, 5 or 6.
21. An expression vector comprising a polynucleotide which encodes a polypeptide comprising the amino acid sequence shown in SEQ ID NO: 2.
22. The expression vector of claim 21 wherein the polynucleotide consists of SEQ ID NOS: 1, 5 or 6.
23. A host cell comprising an expression vector which encodes a polypeptide comprising the amino acid sequence shown in SEQ ID NO: 2.
24. The host cell of claim 23 wherein the polynucleotide consists of SEQ ID NOS: 1, 5 or 6.
25. A purified polypeptide comprising the amino acid sequence shown in SEQ ID NO: 2.
26. The purified polypeptide of claim 25 which consists of the amino acid sequence shown in SEQ ID NO: 2.
27. A fusion protein comprising a polypeptide having the amino acid sequence shown in SEQ ID NO: 2.
28. A method of producing a polypeptide comprising the amino acid sequence shown in SEQ ID NO: 2, comprising the steps of:
 - culturing a host cell comprising an expression vector which encodes the polypeptide under conditions whereby the polypeptide is expressed; and
29. The method of claim 28 wherein the expression vector comprises SEQ ID NOS: 1, 5 or 6.
30. A method of detecting a coding sequence for a polypeptide comprising the amino acid sequence shown in SEQ ID NO: 2, comprising the steps of:
 - hybridizing a polynucleotide comprising 11 contiguous nucleotides of SEQ ID NOS: 1, 5 or 6 to nucleic acid material of a biological sample, thereby forming a hybridization complex; and
 - detecting the hybridization complex.
31. The method of claim 30 further comprising the step of amplifying the nucleic acid material before the step of hybridizing.
32. A kit for detecting a coding sequence for a polypeptide comprising the amino acid sequence shown in SEQ ID NO: 2, comprising:
 - a polynucleotide comprising 11 contiguous nucleotides of SEQ ID NOS: 1, 5 or 6; and
 - instructions for the method of claim 30.
33. A method of detecting a polypeptide comprising the amino acid sequence shown in SEQ ID NO: 2, comprising the steps of:
 - contacting a biological sample with a reagent that specifically binds to the polypeptide to form a reagent-polypeptide complex; and
 - detecting the reagent-polypeptide complex.

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34. The method of claim 33 wherein the reagent is an antibody.

35. A kit for detecting a polypeptide comprising the amino acid sequence shown in SEQ ID NO: 2, comprising:
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an antibody which specifically binds to the polypeptide; and
instructions for the method of claim 33.

10 36. A method of screening for agents which can modulate the activity of a human elongase HSEL01-like protein, comprising the steps of:
contacting a test compound with a polypeptide comprising an amino acid sequence selected from the group consisting of: (1) amino acid sequences which are at least about 50% identical to the amino acid sequence shown in SEQ ID NO: 2 and (2) the amino acid sequence shown in SEQ ID NO: 2; and
15 detecting binding of the test compound to the polypeptide, wherein a test compound which binds to the polypeptide is identified as a potential agent for regulating activity of the human elongase HSEL01-like protein.

20 37. The method of claim 36 wherein the step of contacting is in a cell.

38. The method of claim 36 wherein the step of contacting is *in vitro*.

25 39. The method of claim 36 wherein the step of contacting is in a cell-free system.

40. The method of claim 36 wherein the polypeptide comprises a detectable label.
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41. The method of claim 36 wherein the test compound comprises a detectable label.

42. The method of claim 36 wherein the test compound displaces a labeled ligand which is bound to the polypeptide.
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43. The method of claim 36 wherein the polypeptide is bound to a solid support.

44. The method of claim 36 wherein the test compound is bound to a solid support.

10 45. A method of screening for agents which modulate an activity of a human elongase HSEL01-like protein, comprising the steps of:
15 contacting a test compound with a polypeptide comprising an amino acid sequence selected from the group consisting of: (1) amino acid sequences which are at least about 50% identical to the amino acid sequence shown in SEQ ID NO: 2 and (2) the amino acid sequence shown in SEQ ID NO: 2; and
detecting an activity of the polypeptide, wherein a test compound which increases the activity of the polypeptide is identified as a potential agent for increasing the activity of the human elongase HSEL01-like protein, and
20 wherein a test compound which decreases the activity of the polypeptide is identified as a potential agent for decreasing the activity of the human elongase HSEL01-like protein.
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46. The method of claim 45 wherein the step of contacting is in a cell.

47. The method of claim 45 wherein the cell is *in vitro*.
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48. The method of claim 45 wherein the step of contacting is in a cell-free system.

49. A method of screening for agents which modulate an activity of a human elongase HSEL01-like protein, comprising the steps of:

5 contacting a test compound with a product encoded by a polynucleotide which comprises the nucleotide sequence shown in SEQ ID NOS: 1, 5 or 6; and

10 detecting binding of the test compound to the product, wherein a test compound which binds to the product is identified as a potential agent for regulating the activity of the human elongase HSEL01-like protein.

50. The method of claim 49 wherein the product is a polypeptide.

15 51. The method of claim 49 wherein the product is RNA.

52. A method of reducing activity of a human elongase HSEL01-like protein, comprising the step of:

20 contacting a cell with a reagent which specifically binds to a product encoded by a polynucleotide comprising the nucleotide sequence shown in SEQ ID NOS: 1, 5 or 6, whereby the activity of a human elongase HSEL01-like protein is reduced.

25 53. The method of claim 52 wherein the product is a polypeptide.

54. The method of claim 53 wherein the reagent is an antibody.

30 55. The method of claim 52 wherein the product is RNA.

56. The method of claim 55 wherein the reagent is an antisense oligonucleotide.

57. The method of claim 56 wherein the reagent is a ribozyme.

5 58. The method of claim 52 wherein the cell is *in vitro*.

59. The method of claim 52 wherein the cell is *in vivo*.

60. A pharmaceutical composition, comprising:

10 a reagent which specifically binds to a polypeptide comprising the amino acid sequence shown in SEQ ID NOS: 2; and

a pharmaceutically acceptable carrier.

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61. The pharmaceutical composition of claim 60 wherein the reagent is an antibody.

62. A pharmaceutical composition, comprising:

20 a reagent which specifically binds to a product of a polynucleotide comprising the nucleotide sequence shown in SEQ ID NOS: 1, 5 or 6, and

a pharmaceutically acceptable carrier.

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63. The pharmaceutical composition of claim 62 wherein the reagent is a ribozyme.

64. The pharmaceutical composition of claim 62 wherein the reagent is an antisense oligonucleotide.

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65. The pharmaceutical composition of claim 62 wherein the reagent is an anti-
body.

66. A pharmaceutical composition, comprising:
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an expression vector encoding a polypeptide comprising the amino acid
sequence shown in SEQ ID NO: 2; and
a pharmaceutically acceptable carrier.

67. The pharmaceutical composition of claim 66 wherein the expression vector
comprises SEQ ID NOS: 1, 5 or 6.

68. A method of treating a elongase HSELO1-like protein dysfunction related
disease, wherein the disease is selected from cancer, diabetes, CNS disorder,
asthma, metabolic disease or COPD comprising the step of:
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administering to a patient in need thereof a therapeutically effective dose of a
reagent that modulates a function of a human elongase HSELO1-like protein,
whereby symptoms of the elongase HSELO1-like protein dysfunction related
disease are ameliorated.

69. The method of claim 68 wherein the reagent is identified by the method of
claim 36.
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70. The method of claim 68 wherein the reagent is identified by the method of
claim 45.
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71. The method of claim 68 wherein the reagent is identified by the method of
claim 49.
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Fig. 7
 MDTEMMNSFRGLKIDMOPYDPEFTFODLRFPLIEWVVSFPLIVVY
 MLLILVGGSTYMRTHTRKSPFLQPLIWSPLAFLSPLGCTLRMKENATMVFYVOLKTCV
 PAITDDAVRFWSLFLSKVBEQDAPFLRKLFLFVWYHHSITLFLPSFGXN
 KVPGGEGFMNTFNGTHSTMYTYTMKAALKEPNPLPVITSLQLQMVFGLTIFGLNLY
 IWRQEKGCHTTBFWFSPMLYGTYFLPAHFFRAYLARPKGDASKSQ

Fig. 8
 MEAVVNLVHELMKHADPRIOSYPMSPLISLILITTYTFLS
 LGSPRMANKPQFLDQVLSVYVMTLSPVYVYETMMSWNLSTYTRCDPIDSNSP
 EALRVMYRAVFLMSEKTVLMDOTVFLPLRKDQONTFLHVHSVPLPSWNSWWKJIKIA
 GONGSGFRAMINSVVRVWVLYTGSLAUGPVAQYLWKGKMTAQIQLQFPLVSLHISQ
 YTFMPSDNQYQPLIHLINNYGTIPFLPSNFVHYSTXKLRPRAVQNGCAPATTKV
 KRN

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Fig. 10
 matr̄stvā avadḡfglein əsəscflk̄h vp̄siendfḡi clwp̄fslkf̄v eyFḡpseq
 feflunktf̄l amḡnavali ivȳlliifgg qalrainas hñrlt̄sisi
 v̄lwlmleq̄l vpm̄ynḡlf wsickefaa pklv̄tlyln Ylcktlyln btlv̄lrrk̄
 llflftyhḡ abalcyrḡl igreewvn illhlḡhv̄ myyyfslac ḡirwkwgvw
 tridḡvkyf̄l dkkv̄tlyfasy t̄vayaḡv̄ lprktctgy tqaaaygv̄ lltbyllf̄
 tsidḡvkyf̄k dkktv̄kkesk̄ v̄sayaasf̄ b̄erkarp̄ḡ 1

כְּבָנֵנָה
בְּגַעֲגָגָה
אֲכַבָּבָה
כְּכַבְּבָה
אֲכַבְּבָה
בְּאַכְּבָה
כְּכַבְּבָה
אֲכַבְּבָה
בְּאַכְּבָה
כְּכַבְּבָה
אֲכַבְּבָה

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Fig. 11

BLASTP - alignment of 397_protein against trembl|AF292387|AF292387_1
 product: "CIG30"; Homo sapiens CIG30 gene, partial cds.
 //:9P(AF292387|104444145 product: "CIG30"; Homo sapiens CIG30 gene,
 partial cds.

This hit is scoring at : 5e-139 (expectation value)
 Alignment length (overlap) : 236
 Identities : 100 %

Scoring matrix : BLOSUM62 (used to infer consensus pattern)
 Database searched : nrdb_1_-;

Q:

ATSFPIALIVLIAVGQNYNKKERKGKFNLQGPILWSCLAIFSILGAVRMGIMGVLL

ATSFPIALIVLIAVGQNYNKKERKGKFNLQGPILWSCLAIFSILGAVRMGIMGVLL

H: 1 ATSFPIALIVLIAVGQNYNKKERKGKFNLQGPILWSCLAIFSILGAVRMGIMGVLL

ATSFPIALIVLIAVGQNYNKKERKGKFNLQGPILWSCLAIFSILGAVRMGIMGVLL

TGGLKQTVCPINFIDNSTVKEFWVFLSKVIELEDTAPIILRKPLIFTHWYHHSIVLV

TGGLKQTVCPINFIDNSTVKEFWVFLSKVIELEDTAPIILRKPLIFTHWYHHSIVLV

TGGLKQTVCPINFIDNSTVKEFWVFLSKVIELEDTAPIILRKPLIFTHWYHHSIVLV

YTSFGYKVKYPAAGGMEVTMNGVRAIMVTTKLAANVKPDKMLPLITSLQLQMFVGA

YTSFGYKVKYPAAGGMEVTMNGVRAIMVTTKLAANVKPDKMLPLITSLQLQMFVGA

YTSFGYKVKYPAAGGMEVTMNGVRAIMVTTKLAANVKPDKMLPLITSLQLQMFVGA

IVSILTYIWRDQGCHTTMEHLFWF8TLMITYILPAHFFCQTYIRPKVAKTKSQ

IVSILTYIWRDQGCHTTMEHLFWF8TLMITYILPAHFFCQTYIRPKVAKTKSQ

IVSILTYIWRDQGCHTTMEHLFWF8TLMITYILPAHFFCQTYIRPKVAKTKSQ

236

Fig. 12

BLASTP - alignment of 397_protein against trembl|AF054504|
 gene: "Cig10"; product: "membrane glycoprotein"; Mus musculus
 membrane glycoprotein CIG30; partial cds.; "Cig10"; product: "membrane
 glycoprotein CIG30"; Mus musculus membrane glycoprotein CIG30 (CIG30)
 mRNA; complete cds.; /:gp AF054504|5762267 gene: "Cig10"; product:
 "membrane glycoprotein";
 Mus musculus membrane glycoprotein (CIG30) gene, complete cds.
 CIG30; Mus musculus membrane glycoprotein CIG30 (CIG30) mRNA;
 complete cds.

This hit is scoring at : 5e-113 (expectation value)

Alignment length (overlap) : 271

Identities : 69 %

Scoring matrix : BLOSUM62 (used to infer consensus pattern)

Database searched : nrdb_1_-;

Q: 1 MVTAMNNVSHEVNO-
 LFQPYNPFLSDMRPPEEEWATSFPIALIVLIAVGQNYMWERK

H: 1 M T M N S . . . L Q P Y F E P . D I R P P E Y W . I S P

I.:YL;LJ;VQ;YM;.RK

H: 1 MDTISANPSRGILNDLNPQPYDFETQDLRPFELREYWWSSPLIVVVYLLIVVGCYTMRTRK

GFLNQGPPLWVWFCPLAIFSILGAVRMGIMGVLLQTKVCFINPIDSIVKTVENSWV
 .P:LQ PLIWFSP LAIFSLD: RWW .M.TV:T GLKQTIVCP.

:D:1..V;PWS:1

SFSLSQPLWLSPPFLAIFSILGAVRMGIMGVLLQTKVCFINPIDSIVKTVENSWV

FLLSKVIELEDTAPIILRKPLIFTHWYHHSIVLV
 PLSKV: ELDQTAFLIRKPLIF: HMYHHSITL: TSPYKRNVP: GGWP: TMFGVH:

PLISRVVEFLGDTAFLIRKPLIFTHWYHHSIVLV

IMTYTYTLKAANKVKPDKMLPLITSLQLQMFVGAIVSILTYIWRDQGCHTTTEHPPWS

:MYYTYLKAANKVKPDKMLPLITSLQLQMFVGAIVSILTYIWRDQGCHTTTEHPPWS
 P:LPN:TPFLPAPHPCQTYIRPKVAKTKSQ 270
 P:LY TPFLPAPHPCQTYIRPKVAKTKSQ .Y:RKK K..KSQ
 FMLIGTYTLPAHPPFRAYLRKQKVAKTKSQ 271

614

Fig. 13

BLASTP - Alignment of 397 protein against trembl|AFI170907|AFI170907_1

Gene: "SSC1"; product: "SSC1"; Mus musculus SSC1 (SSC1) mRNA, complete cds. //:SP|AFI170907|8101519 Gen: "SSC1"; product: "SSC1" Mus musculus SSC1 (SSC1) mRNA, complete cds.

This hit is scoring at : 2e-17 (expectation value)

Alignment length (overlap) : 262

Identities : 28 %

Scoring matrix : BLOSUM62 (used to infer consensus pattern)

Database searched : nrdb_1 ;

Q: 21 ELSKDMPPFPEBYWATSPFDIALYLVL-----
IAVQNYMKEKGKFLNQPLIUTWSFCI
EL.K. P .:Y ... P.: . :L
.I.:F.L
H. 10
TELMKHADPQTOSYPAMGSPLLTSALITYYPFDLSPRIMANRKPFOLRGFMIVYNFSL

AIFSLGAVRVMGIMGTVLTLGKL-----
QTVCPCPINIDSTYKFNSWFLPSKTYL
I. S.: . I.: L:G L. . . : P N : . : .

:W.P:LSKVTEL
VILSLY-----
IIVEPLMSGMISITWRCDPFNSNSPEALRMVRVALPMUSKTYTEL

GDTAFTILRK--PLIFIRWHTHSTVLTWTSFGYKNUVPGAG--
WFPTMNEGVHMT
DT...ILRK. .:F.H :HHS : S : K...GG : . .N
.YH. i.M.

MDTVIPILRKDGQVTFLLRVFPHS--
VLPWSWWMGKIAQGNGSFPIAMINSVSHVVMV,
YTYLKAAN--
VVKPPKMLPLMLTQIQLQMEYGAISLILYIMRQDQGCHTMELPWFPI
YY L.A. :P . . :T:Q:O: . : ! Y.. . . : H.
YYCALSALGPVQPYLWWKCHMATAOLIQFVLSLHSIQYYTMMPSCNYQOPILIH
--

LWMT-XPLPAHFFCQTYTRPK 262
.Y. T :PLIPAHFFCQTYTRPK
MGTIPFILFLNFNPYHSTGKCK 259

二

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FIG. 18 (continued)

AF054504-1
 L+LFTSPFGYKVNKPGSGWPTMFGVHSVNTYTYTMQAKLKHEPNLLPMLV
 L++TSGFYKVKVP+GGMP+TMFGVH++MTTYT+KAA+K P
 -LPM+

VVITTSFGYKVNKPGAGWPTMFGVHSVNTYTYTMQAKLKHEPNLLPMLV
 ALI60011
 -22453
 gtaatgtaaatccgggttcaatggcqataattacaggagaccac
 tacgtgaaaatccgggttcatgtacttacaactaccataccattctt

卷之三

AF054504_1 203
 TSIQILOMIVGAIISILTYIWRODGCHTTRHFFNSPMLXGTIFYFILF
 ITSLQIOM +G I IL YIWRQ++QCHTT EH+PMSF+LY
 TSIQILOMIVGAIISILTYIWRODGCHTTRHFFNSPMLXGTIFYFILF
 ALI6001 -2236
 aaaccatcgaggaaacatacggctcaaaatgtttataattact
 tccggatcttttttttttttttttttttttttttttttttttttttt
 cccggggggtaaaaaaaaaaaaaaaaaaaaaaaaaaaaaaaaaaaaa

| | | |
|------------|--------|--|
| AF054504_1 | 252 | AHPFRAVYLPRKGKVASKSQ AHPF + YPRK + KSQ |
| ALI60011 | -22159 | gctttccataaaaggaaaaac cattggatcatccatcgca ccccccggcgacccgcgg |

/ AII 60011. [24800 : 22098]. 6P. Lr
MVT MAMMYSHBUNQFOPNFELSKDMRPFEPYATSFPIALYLVIAQONYMKRKIG
PLNQGQPLJLNSFCALIPLTLLGAVRNUVQKOTCPINFIDNTVKPW93W
LUSRFLDGTAFILRKLPIFMWHSTLVATSPYKVKPAGVSIUTYRDOGCITMEHJFWSP

FIG. 18 (continued)

| | | | |
|----------|--------|-------|-------|
| GeneWise | match | 22098 | 24800 |
| GeneWise | cds | 24199 | 24698 |
| GeneWise | intron | 24399 | 23724 |
| GeneWise | cds | 23722 | 23722 |
| GeneWise | intron | 23931 | 22933 |
| GeneWise | cds | 22231 | 22779 |
| GeneWise | intron | 22780 | 22525 |
| GeneWise | cde | 22232 | 22098 |

FIG. 19

| | |
|---------|-----|
| 1 | 32 |
| outside | 51 |
| TMhelix | 33 |
| inside | 63 |
| TMhelix | 64 |
| outside | 82 |
| TMhelix | 83 |
| inside | 108 |
| TMhelix | 86 |
| inside | 141 |
| TMhelix | 142 |
| outside | 160 |
| TMhelix | 161 |
| inside | 168 |
| TMhelix | 169 |
| inside | 187 |
| TMhelix | 188 |
| inside | 199 |
| TMhelix | 200 |
| outside | 222 |
| TMhelix | 223 |
| inside | 236 |
| TMhelix | 237 |
| inside | 259 |
| TMhelix | 260 |
| inside | 269 |

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- 1 -

Fig. 20

SEQUENCE LISTING

BLASTP - alignment of 397 protein against tremblnew|AF277094|AF277094_1 product: "ELOVL4"; Homo sapiens ELOVL4 mRNA, complete cds. //tremblnew|AF279654|AF279654_1 product: "ELOVL4"; Homo sapiens ELOVL4 gene, exon 6 and complete cds. //gbnew|AF279654|12044051 product: "ELOVL4"; Homo sapiens ELOVL4 gene, exon 6 and complete cds. //gbnew|AF277094|12044043 product: "ELOVL4"; Homo sapiens ELOVL4 mRNA, complete cds.

This hit is scoring at : 2e-15 (expectation value)

Alignment length (overlap) : 242

Identities : 28 %

Scoring matrix : BLOSUM62 (used to infer consensus pattern)

Database searched : nrdb_1_;

Q: 28 PFFEEYWATSFPIALYLVLLAVGQNYMKERKGFLNLQGPLILWSFCLAFSTLGA VRMWG P... W.T. I... YL... G... M.K.R. F... L... F... ::.. H: 41 PIMQSPWPPI- LSISITLYLFVWLGPKW/MKDRPFQMRLVLYNFGMVLNNLFRELF-

IMGTIVL-

TGGLKQTVCFINFDNSTVKFWSWVFLSKVIELGDTAFLILRK--PLIFI MG: . . . Q.V: N: . . . W: .SK: E: DT.F.I.LRK: .F:

MGSYNAGYSYICQSVDYSNNVHEVRRAALWWYFVSKGVVEYLDTVFLRK KNNQVSFL

HWTIHFSTVLYTTSFGYKKNKVPAGGW- VTMNFGVHADMYYTYTTLKAANVKKPKML--PML H YYH,T... .GK ...F..N 'HMY:YY.L.A. .KL ...

IHVHHCTMFTLWwigkWVAGGQAFFGAQLNSFHIVMYSYYGLTAFGPWII KYLWWKRY TISLQIQLQMFVGAIISLTYTWRQDQGCHTIMEHLFWFSFL YMTHF- LEAHFFCOTYR T.LQ:Q.V: .L... .C. .:W.I.Y. .FILE.F.:TY . LTMLQJQTHVTIGHTHALSY--TDC- PFPKWMHWAIIAYAISFLFLNFYTRTYKE

PK 262
PK 270
PK 274

<400> 2
<211> 270
<212> PRT
<213> Homo sapiens

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Met Val Thr Ala Met Asn Val Ser His Glu Val Asn Gln Leu Phe Gln
1 5 10 15

<110> Bayer AG
<120> REGULATION OF HUMAN ELONGASE NSP101-LIKE PROTEIN
<130> Lic079
<150> USEN 60/267,135
<151> 2001-02-08
<160> 10
<170> PatentIn version 3.1
<210> 1
<211> 813
<212> DNA
<213> Homo sapiens
<400> 1
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gatgtgtca aggccatcgag gcccttttc gagggattt ggccaaacctt atcccccata
ggccgtgtatc accttgttttcatcggtgt gggcgaactt acatggaggaa acggaaaggcc
ttcaacatgc aaggccatctt cacccttggg tcctttgtcc tgccatattt cttttatcccg
ggggcagtga ggatgtgggg cattatgggg actgtgtctac ttacccgggg cttttaaaaggaa
accgtgtgtct tcatacaactt catcgataat tccatcgtaa aattttggcc atgggtcttt
tttcatcgca aggccataga actcgaggac acatggccca tcatactggc ttagccggccaa
cttcatcttta ttcaatggta ccaccacagg acatgtgtcg tggtacacaatg ctttggatcc
sagaaaaagg tgccgtgggg aggtgttttc gtccatcgatgg atttttgggtt tcatacgatcc
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cttcatccca ggccgtgtatc ttccgtggatg ctttttttttgg cttatgtcg ctatccatgg
tacatctggg aggggatca gggtgtccac accatgtgg accatgtttt atgggtctcc
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<211> 270
<212> PRT
<213> Homo sapiens

2

3

Pro Tyr Asn Phe Glu Leu Ser Lys Asp Met Arg Pro Phe Phe Cys Gln
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 Tyr Trp Ala Thr Ser Phe Pro Ile Ala Leu Ile Tyr Leu Val Ile
 35 40 45
 Ala Val Gly Gln Asn Tyr Met Lys Glu Arg Lys Gly Phe Asn Leu Gln
 50 55 60
 Gly Pro Leu Ile Leu Trp Ser Phe Cys Leu Ala Ile Phe Ser Ile Leu
 65 70 75 80
 Gly Ala Val Arg Met Trp Gly Ile Met Gly Thr Val Leu Leu Thr Gly
 85 90 95
 Gly Leu Lys Gln Thr Val Cys Phe Ile Asn Phe Ile Asp Asn Ser Thr
 100 105 110
 Val Lys Phe Trp Ser Trp Val Phe Leu Leu Ser Lys Val Ile Glu Leu
 115 120 125
 Gly Asp Thr Ala Phe Ile Ile Leu Arg Lys Arg Pro Leu Ile Phe Ile
 130 135 140
 His Trp Tyr His His Ser Thr Val Leu Val Tyr Thr Ser Phe Gly Tyr
 145 150 155 160
 Lys Asn Lys Val Pro Ala Gly Gly Trp Phe Val Thr Met Asn Phe Gly
 165 170 175
 Val His Ala Ile Met Tyr Thr Tyr Tyr Thr Leu Lys Ala Ala Asn Val
 180 185 190
 Lys Pro Pro Lys Met Leu Pro Met Leu Ile Thr Ser Leu Gln Ile Leu
 195 200 205
 Gln Met Phe Val Gly Ala Ile Val Ser Ile Leu Thr Tyr Ile Trp Arg
 210 215 220
 Gln Asp Gln Gly Cys His Thr Thr Met Glu His Leu Phe Trp Ser Phe
 225 230 235 240

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| | | |
|---|------|---|
| atgtacactt actacactct gaaaggctgcc aacgtgaagc ccccccacat gctggccatg | 600 | Val Val Arg Phe Trp Ser Phe Leu Phe Leu Leu Ser Lys Val Val Glu |
| ctcatcacca gcctgcagat ctgcagat ttgttagag ccatcgtcag catctcaacc | 650 | 115 120 125 |
| tatcatctggaa ggccggatca gggatccaa accacatgg aacactttt ctggtccttc | 720 | Leu Glu Asp Thr Ala Phe Ile Ile Leu Arg Lys Arg Pro Leu Ile Phe |
| atcttgtata tgaccttatt catctctttt gcccacttct tctgcgcagac ctacatcagg | 780 | 130 135 140 |
| cccaaggatca aagccaaagcc caagggccag tgaagggttg gaggasacaa tgaagatcca | 840 | Val His Trp Tyr His Ser Thr Val Leu Leu Phe Thr Ser Phe Gly |
| ggcttcctctt tctccatggcc accaaaggcc tgggtttagtt ttggagaat Gattaggtttgg | 900 | 145 150 155 160 |
| ccttacccctgc atggtttccc cagaggatgt gtgcgcceag gtggctggaa tttttgcag | 960 | Tyr Lys Ala Lys Val Pro Ser Gly Gly Trp Phe Met Thr Met Asn Phe |
| acaagaagg tgaccttggg atgggggtgt ggttgttac tttaatgttt ctgtttttaa | 1020 | 165 170 175 |
| tgtgagggcc aaggaggccc tggatggga gttggggggaa gggggccct aagggtgtat | 1080 | Gly Val His Ser Val Met Tyr Thr Tyr Met Lys Ala Ala Lys |
| tatataattt ctatcccgaa atcttttttc ttatgtctt tttttttaa attaaaggatt | 1140 | 180 185 190 |
| tcaacaa | 1147 | Leu Lys His Pro Asn Leu Leu Pro Met Val Ile Thr Ser Leu Gln Ile |
| <210> 7 | 195 | 200 205 |
| <211> 271 | 210 | Leu Glu Met Val Leu Gly Thr Ile Phe Gly Ile Leu Asn Tyr Ile Trp |
| <212> PRT | 215 | 220 |
| <213> Homo sapiens | 220 | Arg Gln Glu Lys Gly Cys His Thr Thr Glu His Phe Phe Trp Ser |
| <400> 7 | 225 | 230 235 240 |
| Met Asp Thr Ser Met Asn Phe Ser Arg Gly Leu Lys Met Asp Leu Met | 1 | Phe Met Leu Tyr Gly Thr Tyr Phe Ile Leu Phe Ala His Phe His |
| 5 10 15 | 245 | 250 255 |
| Gln Pro Tyr Asp Phe Glu Thr Phe Glu Asp Leu Arg Pro Phe Leu Glu | 20 | Arg Ala Tyr Leu Arg Pro Lys Gly Lys Val Ala Ser Lys Ser Gln |
| 25 30 | 260 | 265 270 |
| Glu Tyr Trp Val Ser Ser Phe Leu Ile Val Val Tyr Leu Leu | 35 | <210> 8 |
| 40 45 | 35 | <211> 279 |
| Ile Val Val Gly Glu Thr Tyr Met Arg Thr Arg Lys Ser Phe Ser Ile | 50 | <212> PRT |
| 55 60 | 50 | <213> Homo sapiens |
| Gln Arg Pro Leu Ile Leu Trp Ser Phe Leu Ala Ile Phe Ser Ile | 65 | <400> 8 |
| 70 75 | 65 | Met Glu Ala Val Val Asn Leu Tyr His Glu Leu Met Lys His Ala Asp |
| 80 | 70 | 1 5 10 15 |
| Leu Gly Thr Leu Arg Met Trp Lys Phe Met Ala Thr Val Met Phe Thr | 85 | Pro Arg Ile Glu Ser Tyr Pro Leu Met Gly Ser Pro Leu Ile Thr |
| 90 95 | 85 | 20 25 30 |
| Val Gly Leu Lys Gln Thr Val Cys Phe Ala Ile Tyr Thr Asp Asp Ala | 100 | 105 110 |
| 105 110 | 95 | 35 40 45 |

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Ile Met Ala Asn Arg Lys Pro Phe Gln Leu Arg Gly Phe Met Ile Val
50 55 60

<210> 9
<211> 292
<212> PRT
<213> Homo sapiens
Tyr Asn Phe Ser Leu Val Ile Leu Ser Leu Tyr Ile Val Tyr Glu Phe
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Leu Met Ser Gly Trp Leu Ser Thr Tyr Thr Trp Arg Cys Asp Pro Ile
85 90 95

Asp Phe Ser Asn Ser Pro Glu Ala Leu Arg Met Val Ala Trp
100 105 110

Leu Phe Met Leu Ser Lys Val Ile Glu Leu Met Asp Thr Val Ile Phe
115 120 125

Ile Leu Arg Lys Lys Asp Gly Gln Val Thr Phe Leu His Val Phe His
130 135 140

His Ser Val Leu Pro Trp Ser Trp Trp Gly Ile Lys Ile Ala Pro
145 150 155 160

Gly Gly Met Gly Ser Phe His Ala Met Ile Asn Ser Ser Val His Val
165 170 175

Val Met Tyr Leu Tyr Tyr Gly Leu Ser Ala Leu Gly Pro Val Ala Gln
180 185 190

Pro Tyr Leu Trp Trp Lys His Met Thr Ala Ile Gln Leu Ile Gln
195 200 205

Phe Val Leu Val Ser Leu His Ile Ser Gln Tyr Tyr Phe Met Pro Ser
210 215 220

Cys Asn Tyr Gln Tyr Pro Ile Ile His Ile Trp Met Tyr Gly
225 230 235 240

Thr Ile Phe Phe Ser Asn Phe Trp Tyr His Ser Tyr Thr
245 250 255

Lys Gly Lys Arg Leu Pro Arg Ala Val Gln Asn Gly Ala Pro Ala
260 265 270

Thr Thr Lys Val Lys Ala Asn
275

Ile Met Ala Asn Arg Lys Pro Phe Gln Leu Arg Gly Phe Met Ile Val
55 60
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<211> 292
<212> PRT
<213> Homo sapiens
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Met Glu Gln Leu Lys Ala Phe Asp Asn Glu Val Asn Ala Phe Leu Asp
1 5 10 15

Asn Met Phe Gly Pro Arg Asp Ser Arg Val Arg Gly Trp Phe Leu Leu
20 25 30

Asp Ser Tyr Leu Pro Thr Phe Ile Leu Thr Ile Thr Tyr Leu Leu Ser
35 40 45

Ile Trp Leu Gly Asn Lys Tyr Met Lys Asn Arg Pro Ala Leu Ser Leu
50 55 60

Arg Gly Ile Ile Leu Thr Leu Tyr Asn Leu Ala Ile Thr Leu Leu Ser Ala
65 70 75 80

Tyr Met Leu Val Glu Leu Ile Leu Ser Ser Trp Glu Gly Gly Tyr Asn
85 90 95

Leu Gln Cys Gln Asn Leu Asp Ser Ala Gly Glu Gly Asp Val Arg Val
100 105 110

Ala Lys Val Leu Trp Trp Tyr Tyr Ser Lys Leu Val Glu Phe Leu
115 120 125

Asp Thr Ile Phe Phe Val Leu Arg Lys Thr Asn Gln Ile Thr Phe
130 135 140

Ile His Val Tyr His His Ala Ser Met Phe Asn Ile Trp Trp Cys Val
145 150 155 160

Leu Asn Trp Ile Pro Cys Gly Gln Ser Phe Gly Pro Thr Leu Asn
165 170 175

Ser Phe Ile His Ile Leu Met Tyr Ser Tyr Tyr Gly Leu Ser Val Phe
180 185 190

Pro Ser Met His Lys Tyr Leu Trp Trp Lys Lys Tyr Leu Thr Gin Ala
195 200 205

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- 10 -

- 11 -

Gln Leu Val Gin Phe Val Leu Thr Ile Thr His Ala Val
210 215 220

Val Lys Pro Cys Gly Phe Pro Phe Gly Cys Leu Ile Phe Gln Ser Ser
225 230 235 240

Tyr Met Met Thr Leu Val Ile Leu Phe Leu Asn Phe Tyr Ile Gin Thr
245 250 255

Tyr Arg Lys Pro Val Lys Lys Glu Leu Gln Glu Lys Glu Val Lys
260 265 270

Asn Gly Phe Pro Lys Ala His Leu Ile Val Ala Asn Gly Met Thr Asp
275 280 285

Lys Lys Ala Gln
290

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<211> 345
<212> PRT
<213> Homo sapiens
<400> 10

Met Asn Thr Thr Thr Ser Thr Val Ile Ala Ala Asp Gln Phe
1 5 10 15

Gln Ser Leu Asn Ser Ser Ser Cys Phe Leu Lys Val His Val Pro
20 25 30

Ser Ile Glu Asn Pro Phe Gly Ile Glu Leu Trp Pro Ile Phe Ser Lys
35 40 45

Val Phe Glu Tyr Phe Ser Gly Tyr Pro Ala Glu Gln Phe Glu Phe Ile
50 55 60

His Asn Lys Thr Phe Leu Ala Asn Gly Tyr His Ala Val Ser Ile Ile
65 70 75 80

Ile Val Tyr Tyr Ile Ile Phe Gly Gly Gln Ala Ile Leu Arg Ala
85 90 95

Leu Asn Ala Ser Pro Leu Lys Phe Lys Leu Leu Phe Glu Ile His Asn
100 105 110

Leu Phe Leu Thr Ser Ile Ser Leu Val Leu Trp Leu Lou Met Leu Glu
115 120 125

Gln Leu Val Pro Met Val Tyr His Asn Gly Leu Phe Trp Ser Ile Cys
130 135 140

Ser Lys Glu Ala Phe Ala Pro Lys Leu Val Thr Leu Tyr Tyr Leu Asn
145 150 155 160

Tyr Leu Thr Lys Phe Val Glu Leu Ile Asp Thr Val Phe Leu Val Leu
165 170 175

Arg Arg Lys Lys Leu Leu Phe Leu His Thr Tyr His His Gly Ala Thr
180 185 190

Ala Leu Leu Cys Tyr Thr Gln Leu Ile Gly Arg Thr Ser Val Glu Trp
195 200 205

Val Val Ile Leu Leu Asn Leu Gly Val His Val Ile Met Tyr Trp Tyr
210 215 220

Tyr Phe Leu Ser Ser Cys Gly Ile Arg Val Trp Trp Lys Gln Trp Val
225 230 235 240

Thr Arg Phe Gln Ile Ile Gln Phe Leu Ile Asp Leu Val Phe Val Tyr
245 250 255

Phe Ala Thr Tyr Thr Phe Tyr Ala His Lys Tyr Leu Asp Gly Ile Leu
260 265 270

Pro Asn Lys Gly Thr Cys Tyr Gly Thr Gln Ala Ala Ala Tyr Gly
275 280 285

Tyr Ile Ile Leu Thr Ser Tyr Leu Leu Phe Ile Ser Phe Tyr Ile
290 295 300

Gln Ser Tyr Lys Lys Gly Gly Lys Lys Val Lys Lys Glu Ser Glu
305 310 315 320

Val Ser Gly Ser Val Ala Ser Gly Ser Ser Thr Gly Val Lys Thr Ser
330 335

Asn Thr Lys Val Ser Arg Lys Ala
340 345

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